

☀ RESEARCH REPORT

Detection of PAH Carcinogens at the Single Molecule Level

V.B. Kartha*, Can C. Ozbal†, Paul L. Skippe†, Steven R. Tannenbaum†, and Ramachandra R. Dasari*

*George R. Harrison Spectroscopy Laboratory, MIT, Cambridge, MA

†Department of Chemistry and Division of Toxicology, MIT, Cambridge, MA

Introduction:

Polycyclic aromatic hydrocarbons (PAH-s) form an important class of chemical carcinogens in view of their ubiquitous nature and assimilation by human beings through environmental pollution, smoking, and food constituents. Benzo[a]pyrene (BaP), is one of a family of PAH-s which undergo metabolic activation *in vivo* and bind covalently to DNA as well as proteins, causing mutations and possibly cancer. In our laboratory we have developed (Dasari RR, 1994, Ozbal CC, 1995) a cryogenic fluorescence spectroscopy technique to quantitatively determine sub-femtomole amounts of BaP adducts of DNA and proteins. We have observed that while synthetic adducts invariably gave a single, strong, symmetric band at 377nm, real-life samples sometimes showed subtle differences in their positions as well as shapes, indicating the presence of possible different types of adducts in different samples. If the nature and source of these differences could be identified it might become possible to get information on the different types of *in vivo* metabolic pathways and different PAH exposure patterns. A suitable method to carry out such characterisation would be to separate and study the individual adducts. Since the total amount of adducts available will frequently be in the subfemtomole range, separation and detection present formidable difficulties. We have therefore set up an HPLC- LIF system for studies involving samples in the picogram range. Some of the initial results are presented here.

Experimental:

An HP-1050 pumping module with an HP-1040A detection system and a Waters C18 (3.9X300mm) column were used for the HPLC separation. A Liconix 3230 He-Cd laser operating at 325nm was used for excitation. The output from the column was passed through a capillary (200 μ m dia.) at flow rates of 0.25-0.5 ml per minute. The laser (10mW) was focused on to the capillary and fluorescence emission was collected by a 40X (NA 0.75) micro-

scope objective. The fluorescence at 377nm was detected using a Spex-1681 model (0.22m) monochromator and R-928 cooled photomultiplier operating at 900-1000 volts. A solution of benzo[a]pyrene derivatives was used to test the performance of the system.

continued on page 8

☀ IN THIS ISSUE ...

- ☀ Personalities
- ☀ Research Report:
The Thermoelastic Basis of Ablation of Biological Tissue, page 2.
- ☀ Spring Seminar Series:
Modern Optics and Spectroscopy
- ☀ May 2 Workshop:
Optical Probes in Biology and Medicine
- ☀ Spectroscopy Laboratory Publications

✱ RESEARCH REPORT

The Thermoelastic Basis of Ablation of Biological Tissue

Douglas Albagli, Marta Dark, Lev Perelman, Charles von Rosenberg, Irving Itzkan, and Michael S. Feld
George R. Harrison Spectroscopy Laboratory, MIT, Cambridge, MA

Laser radiation is currently used in several fields of medicine including ophthalmology, dermatology, urology, and cardiology. However, the interaction between laser light and biological tissue is not completely understood. The fundamental mechanisms of laser ablation are under current debate. In order to optimize laser parameters (wavelength, pulse length, fluence), an understanding of the ablation process is needed. In this article we present our latest results regarding the mechanism of short pulse laser ablation.

A review of experimental results reported in the literature reveals that the energy density required to initiate ablation of biological tissue with nanosecond laser pulses is tenfold less than that required for vaporization. This holds for a wide range of laser wavelengths [Albagli et al, 1994a]. An explanation for this discrepancy is that vaporization (a photothermal process) does not occur at all, and the material ruptures when laser-induced stresses and stress gradients exceed the material's strength. A one dimensional

photomechanical model of laser-induced spallation correctly predicts the reduced energy density, but also predicts that damage should occur approximately one absorption depth beneath the surface [Dingus et al, 1991]. In fact, ablation occurs at or near the surface. For lasers and wavelengths used in ablating biological tissue, the optical absorption depth is usually comparable to the transverse laser dimension, and a one dimensional approximation is not appropriate. The discrepancies can be reconciled by including three dimensional effects. We have solved the fully time dependent three dimensional thermoelastic equation of motion, which predicts significant tensile stresses to form on the surface, (precisely where ablation is observed to occur), and also predicts the thermoelastic expansion of the surface as a function of time.

The surface expansion of tissue immediately after short-pulse laser irradiation is related to the thermal gradient which causes internal stresses and mechanical properties of the tissue. Measuring the surface expansion for fluences below ablation threshold reveals information on these parameters. An interferometric technique, with spatial resolution 3 nm and temporal resolution 3 ns, has been developed to monitor the surface of tissue [Schaffer et al, 1995]. Laser induced thermoelastic expansion is measured by a Michelson interferometer which uses a He-Ne probe laser, the sample is the end mirror in one arm, and a rotating corner cube prism is included in the reference arm. By causing the reference arm length to vary at a constant velocity, the system is given a built-in bias which allows the direction as well as the speed to be measured; thus, the surface position as a function of time can be determined. The thermal input pulse is a 7.5 ns, frequency tripled, Q-switched Nd:YAG laser operating at a wavelength of 355 nm.

Using a fully time dependent numerical solution of the thermoelastic wave equation based on the Adams-Bashforth time stepping method [Albagli, 1994], we can predict the surface expansion of a material given the incident laser profile and material constants. In figure 1 this numerically predicted surface expansion is shown as a function of time for a rounded top-hat laser profile and some typical material parameters. From features of this curve, some unknown material parameters, can be inferred using the known beam radius, w [Itzkan et al, 1995]. These parameters are longitudinal and transverse speeds of sound, C_l and C_t , Poisson's ratio, ν , optical penetration depth, D , and the ratio of the expansion coefficient to the heat capacity, α/C_V . Finally, the stresses can be calculated from the now known material properties and the quasi-steady state displacement, F_4 .

The interferometer was first calibrated using glass and acrylic as samples. Experimental results compared well with theoretical predictions. The measured quasi-steady state values are plotted in figure 2 versus fluence for acrylic and glass. The relation between fluence and displacement is linear as expected. Also in this figure is the theoretically predicted value of this linear relationship as

✱ THE SPECTROGRAPH

Published by the George R. Harrison Spectroscopy Laboratory at the Massachusetts Institute of Technology, Cambridge, MA 02139-4307. Comments, suggestions, and inquiries can be directed to the editors.

Editors: Ramachandra R. Dasari and
Farideh Partovi

✱ GEORGE R. HARRISON SPECTROSCOPY LABORATORY

Director: Michael S. Feld
Associate Director for Scientific Coordination:
Jeffrey I. Steinfeld
Associate Director for Project Coordination:
Ramachandra R. Dasari

The Spectroscopy Laboratory houses two laser research resource facilities. The MIT Laser Research Facility, supported by the National Science Foundation, provides shared facilities for core researchers to carry out basic laser research in the physical sciences. The MIT Laser Biomedical Research Center, a National Institutes of Health Biomedical Research Technology Center, is a resource center for laser biomedical studies. The LBRC supports core and collaborative research in technological research and development. In addition, it provides advanced laser instrumentation, along with technical and scientific support, free of charge to university, industrial and medical researchers for publishable research projects. Write or call for further information or to receive our mailings, (617) 253-9774.

Figure 1. The time evolution of the thermoelastic expansion, predicted by the three-dimensional model allows the determination of mechanical, optical, and thermal properties from features in the surface movement. These features also allow determination of the magnitude of the induced stresses. Quasi steady-state displacement is normalized to 100.

Figure 2. Quasi steady-state displacements of acrylic and glass as a function of fluence.

calculated from the manufacturer's data for these materials, with no free parameters. For glass, the agreement is excellent, even to ablation threshold. In acrylic, there is initial agreement at low fluences, but an as yet undetermined non-linear effect causes the displacements to be lower than expected at high fluences. Also, for acrylic there is a larger spread in the data with each subsequent shot at the same location, each one producing less thermoelastic expansion. Small permanent defects, observed with a microscope, are created in acrylic by sub-threshold laser pulses.

The acrylic samples were studied microscopically, and a large number of defects were observed. Under high magnification the defects appear to be fracture patterns. They are concentrated near the surface of the sample, and the number of defects increases with subsequent laser pulses. Defect formation also correlates with the non-linear fall-off of quasi-steady state displacement as a function of fluence shown in figure 2. These "microcracks" weaken the overall strength of the acrylic, and their accumulation may play a significant role in the ablation process, called the "incubation

continued on page 4

effect,” where a material does not seem to be affected by n pulses at a given sub-threshold fluence, but then ablates on the $n+1$ pulse.

An analogous effect to the microcrack in hard tissue is cavitation in soft tissue. The thermoelastic surface expansion of meniscus, a soft biological tissue, showed an unexpected feature; and additional expansion lasting 1-4 μ s. These results are described in reference 3 in detail. Laser fluences used were very low, corresponding to a temperature of $<10^\circ$ C. Water doped with FeCl_3 was also used to determine whether the anomalous behavior in meniscus could be ascribed to water which comprised 70% of meniscal tissue. The same feature which appeared in meniscus was observed in water and can be explained by the creation of cavitation bubbles. The observed times of growth and decay are consistent with the times for formation and collapse of cavitation bubbles. Cavitation is created in water under tension. Since the collapse of cavitation bubbles is known to be destructive to adjacent solid material, the presence of cavitation at such low fluences may have serious consequences for medical laser procedures.

We studied the onset of ablation in beef cortical bone, a tissue whose behavior is similar to that of acrylic. Figure 3 shows typical surface displacement of bone. The quasi-steady state displacement was normalized to 100 for several fluences, and time dependent behavior is identical throughout the fluence range. The theoreti-

cally predicted movement is compared to the measured movement in figure 3. However, the predicted alternating contractions and expansion were “washed out.” For a turbid media such as bone, scattering within the target will change the temperature distribution and blur the sharp temperature gradient at the radial edge of the laser beam. The temperature distribution was calculated numerically using the absorption coefficient and the effective scattering coefficient for in a Monte Carlo calculation [Itzkan et al, 1995]. The predicted thermoelastic surface motion of the bone, using this new temperature distribution, is also plotted in figure 3. The agreement between theory and experiment considerably improved when scattering effects are taken into account, lending credence to the hypothesis that scattering is responsible for the “washing out” of the small expansions and contractions.

In the apparatus, the He-Ne probe beam can be translated across the sample's surface relative to the pump beam, so that it can be positioned at different radii, including radii outside the irradiated area. Even though the material receives no laser irradiation here, it shows displacement since it is pulled up by the “hot” adjacent material. Off-center monitoring allows us to observe the behavior of the rim during ablation. Although the surface is destroyed once ablation occurs and material is ejected, we can monitor the surface outside the irradiated area ($r > w$) to determine when ablation occurs. Figure 4 shows the motion of bone just

Figure 3. Surface movement of cortical bone compared to theoretical predictions which include a correction due to optical scattering. Four different fluences are displayed with quasi steady-state values normalized to 100.

outside the irradiated area when irradiated with sufficient energy to cause ablation, compared to the sub-threshold theoretical curve. Ablation occurs at a time about 350 ns after the laser pulse. There is an abrupt departure from the theoretical curve, the surface at the rim undergoes a large contraction to a position 200 nm below the original surface position, followed by a slow recoil expansion for several microseconds to the limit to which we monitored time in this measurement. This experiment was repeated for acrylic and surface damage was observed under a microscope. The appearance of surface damage always corresponded with the large negative surface motion of the rim.

Interferometric surface monitoring is an important tool for studying the fundamental mechanisms of ablation of biological tissue. A three dimensional theoretical model for thermoelastic surface expansion after sub-ablation threshold laser irradiation has been developed which establishes the relationship of this movement to important mechanical and optical properties of the tissue. The surface movement of bone, glass, and acrylic were measured after irradiation with a 7.5 ns, 355 nm pulse. The results for glass and acrylic were quantitatively consistent with the theoretical model presented; although, acrylic did show a deviation from theory as defects were formed in the material. The results for bone were in good agreement with the theory after the effects of scattering were taken into account. Finally, the interferometric surface monitoring

technique has the ability to monitor the ablation event by placing the probe beam outside of the pump beam area. Currently, further studies are underway to measure the optical and mechanical properties of bone, and to determine the threshold for cavitation and its effects in soft tissue ablation.

References

- Albagli D, Perelman LT, Janes G S, von Rosenberg C, Itzkan I, & Feld MS. *Las Life Sci.* 6, :55-68. (1994a).
- Dingus RS & Scammon RJ in *Laser Ablation, Mechanisms and Applications*, eds. Miller JC & Haglund Jr., RF (Springer-Verlag, New York, NY), 389, :180-190. (1991).
- Schaffer J L, Dark M, Itzkan I, Albagli D, Perelman LT, von Rosenberg C, & Feld M S. *Clin. Orthopaedics.* 310, :30-36. (1995).
- Albagli D "Fundamental Mechanisms of Pulsed Laser Ablation of Biological Tissue", (PhD Thesis, MIT), :159-177. (1994).
- Itzkan I, Albagli D, Dark M, Perelman LT, von Rosenberg C, & Feld M.S. *Proc. of Natl. Acad. of Sciences.* in press (1995). ■

Figure 4. Ablation of bone is monitored by placing the He-Ne probe beam outside of laser irradiated area ($w=400$ mm, $r=700$ mm). The fluence used, 35.4 mJ/mm², is the threshold value for causing ablation. At about 350 ns after the laser pulse, the cold rim recoils to a depth of -250 nm, and then expands. These motions are very large compared to the sub-threshold values and appear only when ablation occurs.

Seminar on

MODERN OPTICS AND SPECTROSCOPY

FALL SEMESTER, 1994

- February 21** Curt Wittig, University of Southern California
Ultraviolet Photochemistry of HI Dimers
- February 28** Mara Prentiss, Harvard University
Neutral Atom Photography
- March 14** Edward A. Hinds, Yale University
Testing Fundamental Symmetries with Molecules
- March 21** Ronald R. Parenti, Lincoln Laboratory, MIT
Adaptive Optics for Astronomy
- April 11** Richard Gottscho, AT&T
Getting into the Display Business (Again)
- April 18** Herbert Walther, Max-Planck-Institut Fur Quantenoptik
Single Atoms in Cavities
- April 25** Lev Perelman, Spectroscopy Laboratory
Photon Migration for Spectroscopy and Imaging
- May 2** David Wineland, National Institute of Standards
Entangled States for Spectroscopy and Computations

May 9	Special Event Fourth Annual Richard C. Lord Lecture Richard Zare, Stanford University "Structure" of the Nitric Oxide Ionization Continuum
--------------	---

TUESDAYS, 11:00-12:00, Marlar Lounge (37-252), Ronald E. McNair Building

Refreshments Served Following the Seminar

Sponsored by George R. Harrison Spectroscopy Laboratory,
Research Laboratory of Electronics, Schools of Science and Engineering,
Plasma Fusion Center and Industrial Liaison Program,
Massachusetts Institute of Technology
Rowland Institute for Science

Lester Wolfe Workshops in Laser Biomedicine

Optical Probes in Biology and Medicine

Tuesday, May 2, 1995, 4:00-7:00 PM

**Can Two-Photon Excitation Illuminate Medical Applications of Laser
Microscopy**

Watt W. Webb, Cornell University

**Photon Migration in Turbid Media With Early Arriving Photons: To-
wards Optical Imaging Through the Human Body**

Jun Wu, MIT Spectroscopy Laboratory

**Resonance Raman and Gd Vibronic Sideband Spectroscopy as Probes
of Protein Structure and Dynamics**

Joel Friedman, Albert Einstein College of Medicine

Applications of Flash-Photolysis to Biological Photochemistry

Robert Redmond, MGH-Wellman Laboratories

HST Auditorium (E25-111), Whitaker College Building, MIT
45 Carlton Street, Cambridge

Refreshments at 3:30 P.M.

**Sponsored by MIT Laser Biomedical Research Center,
MGH Wellman Laboratories, MIT Industrial Liaison Program, &
Harvard-MIT Division of Health Sciences and Technology**

☀ PERSONALITIES

Dr. Steven R. Tannenbaum was born in New York City and began his lifetime association with MIT in 1954, receiving the S.B. degree in 1958 and the Ph.D. degree in 1962. This was followed by appointment to the faculty of the Department of Nutrition and Food Science (later to be renamed the Department of Applied Biological Sciences). Promotions ensued to Associate Professor and then Professor in 1974. He has been Registration and Admissions Officer of the both the Department of Applied Biological Sciences (1982-1988) and more recently of the Division of Toxicology (1988-present). He is also an Associate Director of the MIT Center for Environmental Health Sciences. Other MIT activities have included the Committees on Educational Policy, Academic Performance, and the committee that founded the Independent Activities Period.

Dr. Tannenbaum's re-riety of fields ranging from tional biochemistry to toxicology. Since the mid-1970's his main interests have centered on carcinogen chemistry and biocompounds that are in air, food, and water. Research on the relationships between environmental exposure and human disease endogenous synthesis of carcinogens. In particular, the investigation of the N-nitroso compounds led to studies on the pharmacokinetics of nitrate and nitrite. A major discovery followed from these experiments: nitrate and nitrite were themselves synthesized in mammals de novo from reduced nitro-species (NH_4^+ (NO_3^-), and the biosynthesis was stimulated by infection. These findings contributed to the subsequent discovery by other investigators that the source of nitrate was in fact nitric oxide synthesized by oxida-

It was the research on carcinogens that are found in the environment that led to his current program to develop a method to measure exposure at the sub-cellular level. Efforts with mass spectrometry for a number of different types of molecules, but another dimension was required. This was provided by the current collaboration with Dr. Ramchandra Dasari and his co-workers in the area of LIF. A number of approaches have yielded impressive results in the quantitative analysis of pyrene-containing structures at attomolar levels, and qualitative results on molecular structure using cryogenic laser-induced fluorescence (LIF) line-narrowing spectroscopy. A current project involves construction of a molecular and/or cell separation system that will push the envelope down to the single molecule level of detection. Support for all of this research has come from the National Cancer Institute, the National Institute of Environmental Health Sciences, and the Superfund.

In addition to his activities at MIT, Dr. Tannenbaum is very active at the national and international level of professional societies. He is currently on the editorial boards of four journals and has served on numerous committees of the American Association for Cancer Research. He is also currently a member of the Board of Scientific Counselors of the Division of Cancer Etiology of the National Cancer Institute, and tries to find some time for his two grandchildren, Bailey (5) and Seth (0.5) Hanselman. When not in the vicinity of MIT he can sometimes be found wandering the beaches of Cape Cod.

STEVEN R. TANNENBAUM

search has spanned a variety of fields ranging from food chemistry to nutritional biochemistry to toxicology. Since the mid-1970's his main interests have centered on carcinogen chemistry, particularly food, and water. Research between environmental exposure and human disease endogenous synthesis of carcinogens. In particular, the investigation of the N-nitroso compounds led to studies on the pharmacokinetics of nitrate and nitrite. A major discovery followed from these experiments: nitrate and nitrite were themselves synthesized in mammals de novo from reduced nitro-species (NH_4^+ (NO_3^-), and the biosynthesis was stimulated by infection. These findings contributed to the subsequent discovery by other investigators that the source of nitrate was in fact nitric oxide synthesized by oxida-

the very low levels of carcinogens that led to his current biomarkers of carcinogen exposure at the sub-cellular level. Early efforts with mass spectrometry gave excellent results for a number of different types of molecules, but another dimension was required. This was provided by the current collaboration with Dr. Ramchandra Dasari and his co-workers in the area of LIF. A number of approaches have yielded impressive results in the quantitative analysis of pyrene-containing structures at attomolar levels, and qualitative results on molecular structure using cryogenic laser-in-

■

Results and Discussion:

Figure 1 shows the HPLC detector output and corresponding absorption spectrum for a 2.75 nmole (total) sample. The absorption spectra of all the fractions were identical indicating that the sample contained four different pyrene derivatives. The fluorescence signal from 110 picomole (total) of sample is shown in Figure

2. The inset in Fig.2 corresponds to 11 attomole (total) of the same sample. Figure 2 gives a realistic estimate of the minimum detectable quantity with the present setup, which will be in the zeptomole range. But a more clear picture of the capability of the system can be obtained when we consider the signal output at any

Figure 1. HPLC peaks (bottom) and absorption spectrum (top) of a 2.75 nanomole (total) solution of benzopyrene derivatives. All the four HPLC peaks gave same spectra indicating that the sample contains four different pyrene derivatives only.

instant of time for the 11 attomole sample. The laser beam is focussed to about 10 microns. With a capillary radius of 100 microns the volume of solution observed by the system at any time is about 3×10^{-7} ml. Only about 4 attomoles contribute to the most intense peak, and this amount comes out of the system in about 0.25 ml. (A flow rate of 0.5 ml/minute, with peak half width of 30 seconds). Hence the quantity detected at any time is about $4 \times 3 \times 10^{-7} \times 6 \times 10^5 = 3$ molecules. This is not surprising, since, in one minute the sample travels about 17 meters and the time spent in the laser beam is about 35 microseconds. With life times of the order of 100 nseconds, each molecule thus gets excited several hundred times and emits a large number of photons. It is thus evident that the present system will be capable of detecting single molecules of BP with very little further effort.

An important consequence of the high sensitivity achieved in the present system is the possibility that it can be used to sort out intact cells containing PAH adducts. Cells from smokers are estimated to contain on an average about 1000 PAH molecules per cell, while non-smokers may have less than a few hundred molecules of PAH per cell. It will not be very difficult with improvements in the present system to carry out such sorting, enabling a preconcentration of adducted cells with any desired degree of adduction. The cells can then be examined for the variations in spectral properties, type and nature of adducts etc. as was mentioned earlier.

It should be noted that many single molecule detection experiments (Nie S, 1994, Fan F-RF, 1995) use relatively high concentrations of samples (nM-mM) and look at an extremely small volume (pico-zepto litres). That is, one has a large number of molecules at his disposal but wants to look at only one at a time. While this is necessary for study of dynamics of single molecules, in applications of detection and quantitation, one faces the reverse problem. That is one has only a very small number of molecules available and wants to look at all of them simultaneously. The two areas merge when a specific molecule can be localised at a given point in space and one can study it as long as necessary.

References:

- Dasari RR, Kartha VB, Singh K, Ozbal CC, Skipper PL, and Tannenbaum SR, ICORS-94, Satellite Meeting, China, August 28-30, (1994).
- Ozbal CC, Kartha VB, Skipper PL, Dasari RR, and Tannenbaum SR, *Proceed. American Assoc. Cancer Research*, 36, 134, (1995).
- Nie S, Chiu DT, and Zare RN, *Science*, 266, 1018, (1994).
- Fan F-R F and Bard AJ, *Science*, 267, 871, (1995).

continued on page 12

★ SPECTROSCOPY LABORATORY PUBLICATIONS

“A new cavity ring-down technique and its application to measurement of ultra-slow velocities”, An K, Yang C, Dasari RR, Feld MS. *Optics Letters*, in press (1995).

“An analytical model for evaporative cooling of atoms”, Davis KB, Mewes M-O, and Ketterle W, *Appl. Phys. B*, in press (1995).

“An Atomic-Ion-in-Molecule Analysis of the Low-Lying States of NiO”, Friedman-Hill EJ and Field RW, *J. Mol. Spectrosc.*, in press (1995).

“Approach to single-molecule detection with SERRS”, Kneipp K, Wang Y, Dasari RR, Feld MS, *Appl. Spectrosc.* in press (1995).

“Critical density fluctuations of the cornea”, Matsuura T, Gorti S, Saishin N, Tanaka T, *Proc. NAS*, in press (1995).

“Detection and Characterization of Tissue Lesions with NIR Raman Spectroscopy”, Feld MS, Manoharan R, Salenius J, Orenstein-Candona J, Romer TJ, Brennan JF, Wang Y. *Proceedings of the Biomedical Optics '95, SPIE*, San Jose, CA, February 4-10, in press (1995).

“Detection of Single Molecules with Surface-Enhanced Raman Scattering: A Feasibility Study Using Rhodamine 6G”, Kneipp K, Wang Y, Dasari RR, Feld MS, in press (1995).

“Diagnostic Fluorescence Spectroscopy of Oral Mucosa”, Roy K, Bottril ID, Ingrams DR, Pankratov MM, Rebeiz EE, Woo P, Kabani S, Shapshay SM, Manoharan R, Feld MS. *Proceedings of the Biomedical Optics '95, SPIE*, San Jose, CA, February 4-10, in press (1995).

“Direct Measurements of Rotational and Vibrational Relaxation in Methane Overtone Levels by Time-Resolved Infrared Double-Resonance Spectroscopy”, Klaassen JJ, Coy SL, Steinfeld JJ, and Abel B, *J. Chem. Phys.* in press (1995).

“Dis-Bond Detection and the Possibility of Interfacial Stiffness Measurement with Real-Time Impulsive Stimulated Thermal Scattering”, Rogers JA, Nelson KA, *J. Adhesion*, in press (1995).

“Electrophile-Promoted Carbyne-CO Coupling at a Tantalum Center”, Protasiewicz JD, Bronk BS, Masschelein A, and Lippard SJ, *Organometallics*, in press (1995).

“In situ Histochemical Analysis of Human Coronary Arteriy by Raman Spectroscopy Compared with Biochemical Assay”, Brennan JF, Romer TJ, Wang Y, Tercyak AM, Wang Y, Fitzmaurice M, Lees RS, Dasari RR, Kramer JR, Dasari, Feld MS. *Proceedings of the Biomedical Optics '95, SPIE*, San Jose, CA, February 4-10, in press (1995).

“Laser-Induced Fluorescence Endoscopic Imaging for Detection of Colonic Dysplasia”, Wang TD, Wang Y, Van Dam J, Crawford JM, Preisinger EA and Feld MS. *Proceedings of the Biomedical Optics '95, SPIE*, San Jose, CA, February 4-10, in press (1995).

press (1995).

“Laser-Induced Fluorescence Spectroscopy of Colonic Dysplasia: Prospects for Optical Histological Analysis”, Manoharan R, Zonios G, Cothren RM, Arendt J, Van Dam J and Feld MS. *Proceedings of the Biomedical Optics '95, SPIE*, San Jose, CA, February 4-10, in press (1995).

“Mechanisms of Meniscal Tissue Ablation by Short Pulse Laser Irradiation” Schaffer JL, Dark M, Itzkan I, Albagli D, Perelman L, von Rosenberg, Feld MS. *Clinical Orthopedics*, in press (1995).

“Mechanistic Studies of the Reaction of Dioxygen with Dinuclear Iron(II) Compounds to Form (m-Peroxo)diiron(III) Complexes”, Feig AL and Lippard SJ, *J. Am. Chem. Soc.*, in press (1995).

“Near-Infrared Excitation Profile Study of Surface-Enhanced Hyper-Raman Scattering (SERS) and Surface-Enhanced Raman Scattering (SERS) by Means of Tunable Mode-Locked Ti:Sapphire Laser Excitation”, Kneipp K, Kneipp H, Seifert F. *Chemical Physics Letters*, in press (1995).

“NIR-SERS of neurotransmitters in colloidal silver solution”, Kneipp K, Wang Y, Dasari RR, Feld MS, *Spectrochim. Acta A*, in press (1995).

“One-Step Absolute Frequency Stabilization of a Ti:Sapphire Laser using FM Lamb-Dip Spectroscopy”, An K, Dasari RR, Feld MS. *Applied Physics Letters*, in press (1995).

“Optical Imaging in Turbid Media Using Early Arriving Photons”, Perelman LT, Wu J, Itzkan I, Wang Y, Dasari RR, and Feld MS. *Proceedings of the Biomedical Optics '95, SPIE*, San Jose, CA, February 4-10, in press (1995).

“Origin of Structural Inhomogeneities in Polymer Gels”, Matsuo ES, Sun ST, Li Y, Tanaka T, *Macromolecules*, in press (1995).

“Preparation of II-VI Quantum Dot Composites by Electropray OMCVD”, Danek M, Jensen KF, Murray CB, Bawendi MG, *J. Cryst. Growth*, in press (1995).

”Preparation of II-VI Quantum Dot Composites by Electropray OMCVD”, Danek M, Jensen KF, Murray CB, Bawendi MG, in “Growth, Processing, and Characterization of Semiconductor Heterostructures”, *MRS Vol. 326*, in press (1995).

“Quantum Beat Spectroscopic Studies of Zeeman Anticrossings in the \tilde{A}^1A_u State of the Acetylene Molecule (C_2H_2)”, Dupr'e P, Green PG, and Field RW, *J. Mol. Spectrosc.* in press (1995).

“Rotational, Vibrational, and Orientation Relaxation in Methane and Its Relation to Collision Broadening”, Steinfeld JI, et al., invited presentation at CCP6 Workshop: Inelastic Collision and Dynamics in the Atmosphere”, Durham, U.K. (July 1995).

“Spectral Diagnosis of Human Coronary Artery: A Clinical System for Real Time Analysis”, Kramer JR, Brennan JF, Romer TJ, Wang Y, Dasari RR, Feld MS. *Proceedings of the Biomedical*

Optics '95, SPIE, San Jose, CA, February 4-10, in press, (1995).

“Stark and Zeeman Spectroscopy of Acetylene: Locating a Triplet Isomerization Barrier”, Green PG, Dupr'e P, Lombardi M, Jost R, Kinsey JL, and Field RW, *J. Chem. Phys.* in press (1995).

“Synthesis and Structural Characterization of $[Cr(t-C_4H_9HNCJCNH-t-C_4H_9)(CN-t-C_4H_9)_4]I$; Reductive Coupling of Isocyanide Ligands in a First Row Transition Metal Complex”, Acho JA and Lippard SJ, *Organometallics*, in press (1995).

“Synthesis, Structure, and Electronic Properties of a Mixed-Valent Dodecairon Oxo Complex, a Model for the Biomineralization of Ferritin”, Taft KL, Papaefthymiou GC, and Lippard SJ, *Inorg. Chem.*, in press (1995).

“The Acetylene S_0 Surface: From Dispersed Fluorescence Spectra to Polyads to Dynamics”, Solina SAB, O'Brien JP, Field RW, and Polik WF, *Ber.Bunsen.Phys.Chem.*, in press (1995).

“The Acetylene S_0 Surface: From Dispersed Fluorescence Spectra to Polyads to Dynamics”, Solina SAB, O'Brien JP, Field RW, and Polik WF, *Berichte der Bunsengesellschaft fur Physikalische Chemie* \underbar, in press (1995).

“The Photomechanical Basis of Laser Ablation of Biological Tissue”, Perelman LT, Albagli D, Dark M, Schaffer J, von Rosenberg C, Itzkan I, Feld MS. *Proceedings of the Biomedical Optics '95, SPIE*, San Jose, CA, February 4-10, in press (1995).

“The Thermoelastic Basis of Short Pulsed Laser Ablation of Biological Tissue”, Itzkan I, Albagli D, Dark ML, Perelman LT, von Rosenberg C, Feld MS. *Proceedings of National Academy of Science*, in press (1995).

“The $\Omega=1$ van der Waals and $\Omega=O^+$ Double Well Potentials of $Xe\ 6s + Kr\ ^1S_0$ Determined from Tunable Vacuum Ultraviolet Laser spectroscopy”, Pibel CD, Yamanouchi K, Miyawaki J, Tsuchiya S, Rajaram B, and Field RW, *J. Chem. Phys.* in press (1995).

“Three-Dimensional Imaging of Objects Embedded in Turbid Media with Fluorescence and Raman Spectroscopy”, Wu J, Wang Y, Perelman LT, Itzkan I, Dasari RR, Feld MS. *Applied Optics*, in press (1995).

“Time Dependent Photon Migration using Path Integrals”, Perelman LT, Wu J, Wang Y, Itzkan I, Dasari RR, Feld MS. *Physical Review E*, in press (1995).

“Time-Resolved 3-D Imaging of Fluorescent Objects in Turbid Media”, Wu J, Wang Y, Perelman LT, Itzkan I, Dasari RR, Feld MS. *Proceedings of the EOS/SPIE, conference on Biomedical Optics '94, Lille, France, 2326:252-256*, (1995).

“Time-Resolved Multichannel Imaging of Fluorescent Objects Embedded in Turbid Media”, Wu J, Wang Y, Perelman LT, Itzkan I, Dasari RR, Feld MS. *Optics Letters* in press (1995).

“Vibrational Effects on Rotational Energy Transfer and *Vice Versa*”, Steinfeld JI, to appear in *Vibrational Energy transfer Involving Large and Small Molecules* (Barker, JR, ed.), *Adv. Chem. Kinetics and Dynamics*, 2 (JAI Press), (1995). ■

Figure 2. Fluorescence signal from 110 picomole (total) of sample of Figure 1. Excitation He-Cd laser, 12 mW. Emission 377 nm. Inset shows the fluorescence signal from 11 attomole (total) of the same sample. ■