

*Winter School on Biomolecular Solid State NMR*

*January 20 - 25, 2008*

**“Selected Topics Lecture”**

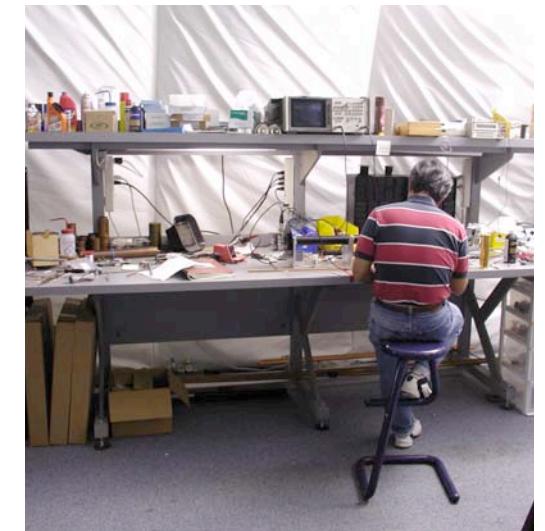
***Stanley J. Opella***



# *University of California, San Diego*



# *Center for NMR Spectroscopy and Imaging of Proteins*



# *Solid-state NMR of membrane proteins in phospholipid bilayers.*

## bilayer samples

150  $\mu\text{l}$  volume

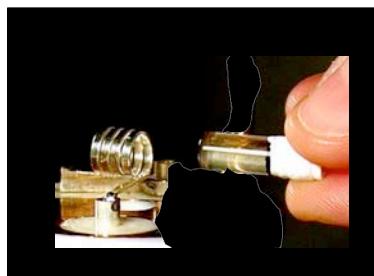
$q = 3.2$  (DMPC:DHPC)

1 mg - 3 mg protein

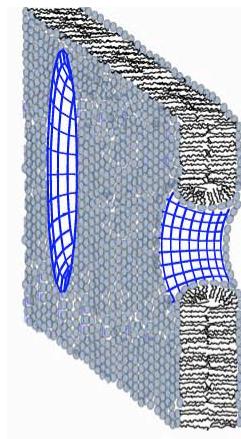
28% lipid w/v

pH = 4 - 8

15 °C – 50 °C



$B_o$



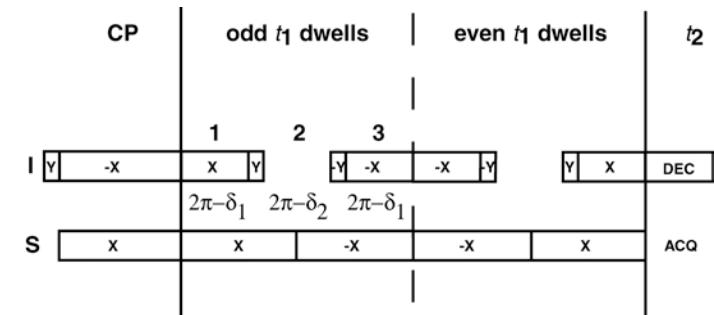
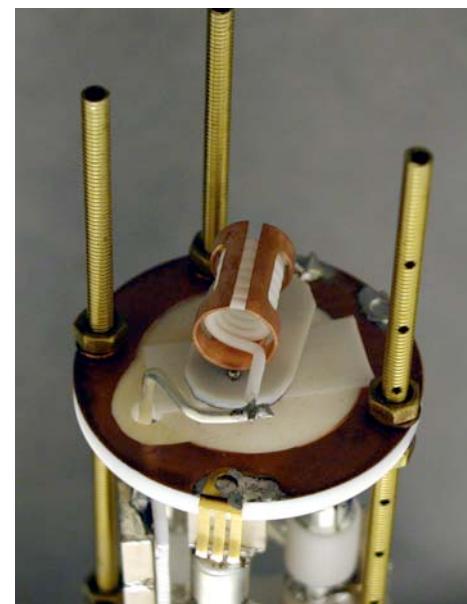
## NMR probes

Outer modified Alderman-Grant

Inner 5 mm solenoid

## pulse sequences

SAMPI4



## *Selected Topics*

- **Part I: Recent developments in solid-state NMR of aligned samples.**
  - Lipid bilayers.
  - Double-resonance experiments.
  - Triple-resonance experiments.
- **Part II: Phospholipid bilayers are essential for biological relevance.**
  - Mercury transport membrane proteins.
  - Vpu from HIV.

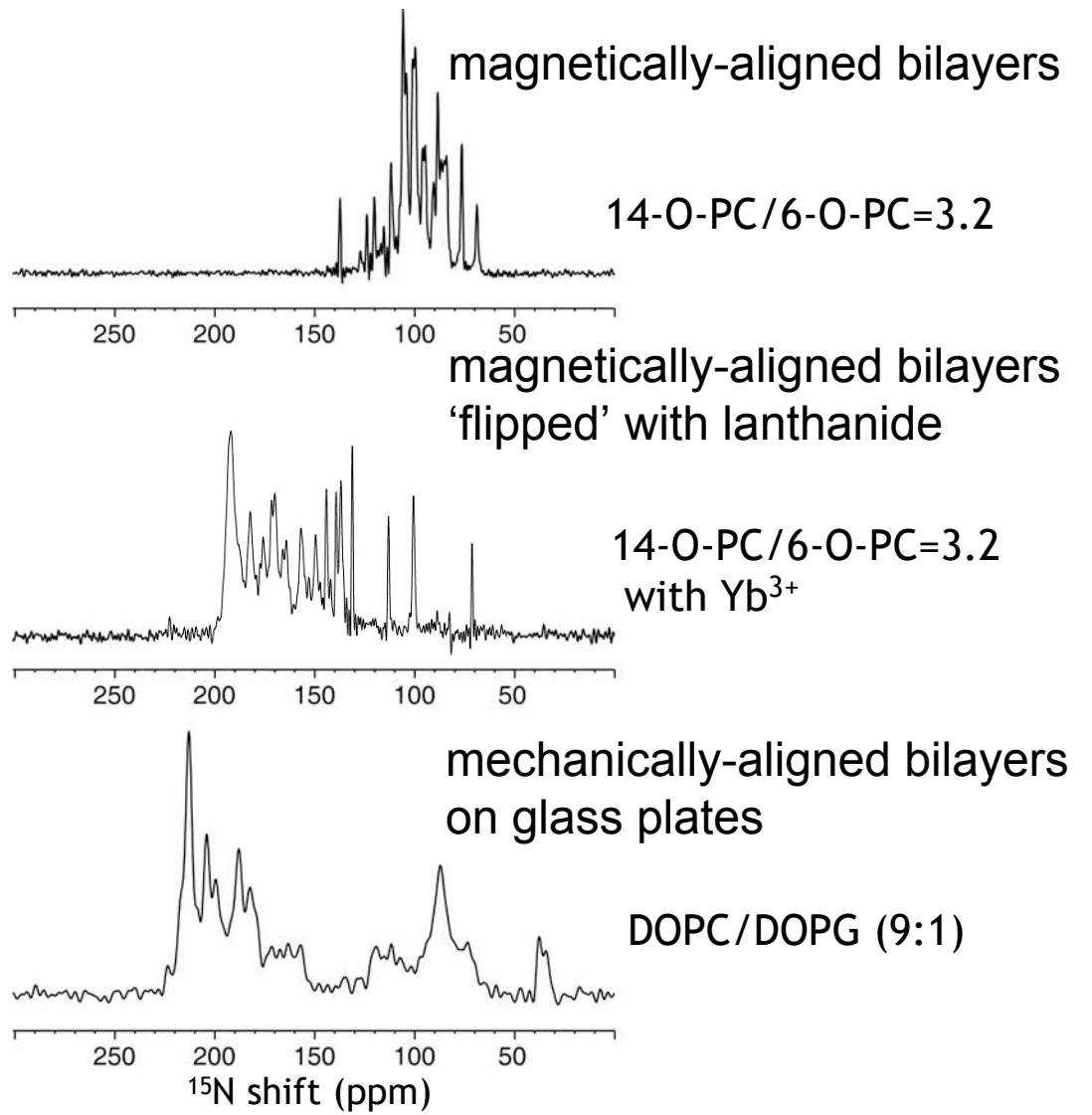
## **Part I: Recent developments in solid-state NMR of aligned samples.**

*Lipid bilayer samples.*

# *Solid-state NMR spectra of Pf1 coat protein in phospholipid bilayers.*



$B_0$



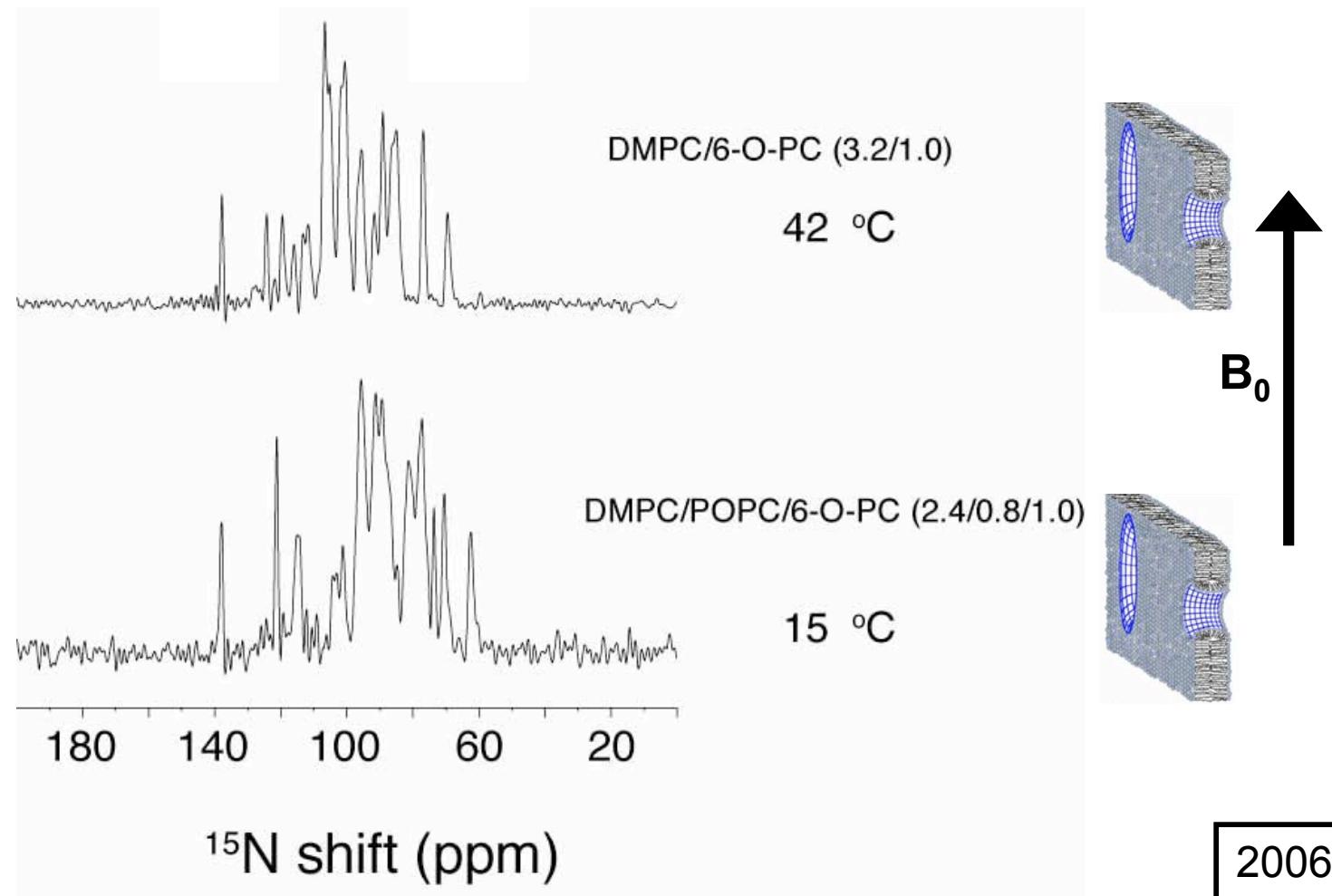
Pf1 coat protein

# *Magnetic alignment of bilayers at low temperatures without loss of resolution.*

## **Effects of Lipid Chain Length and Unsaturation on Bicelles Stability. A Phosphorus NMR Study**

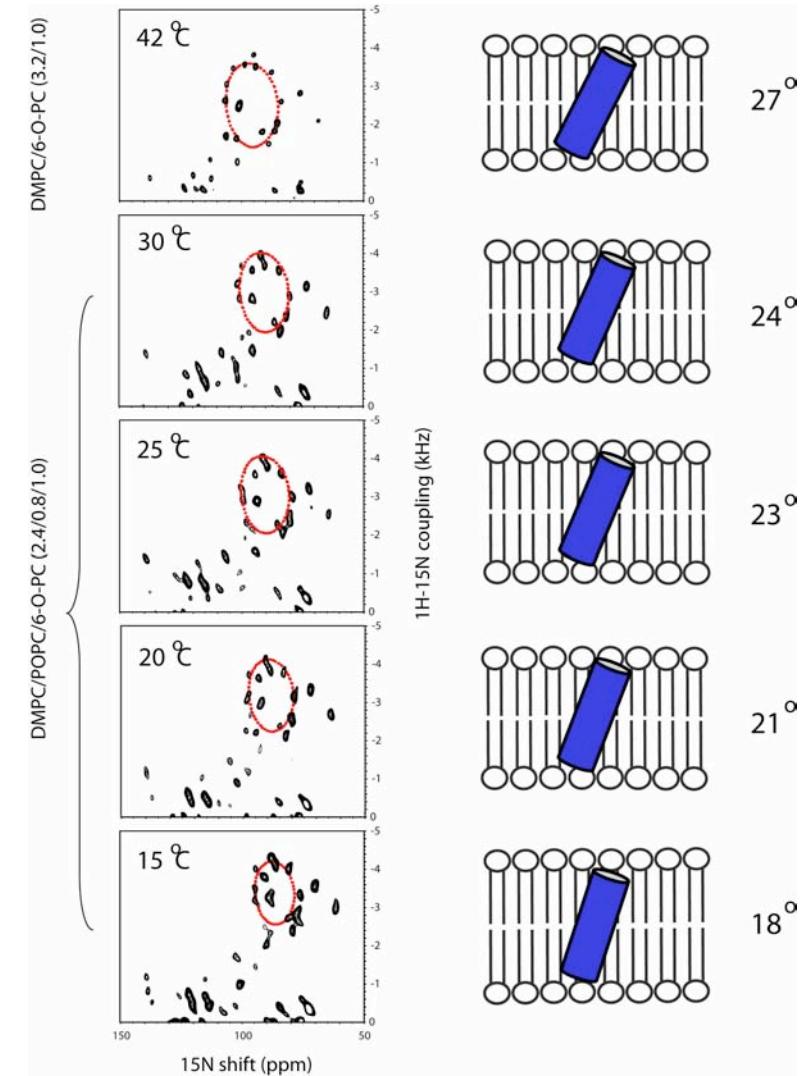
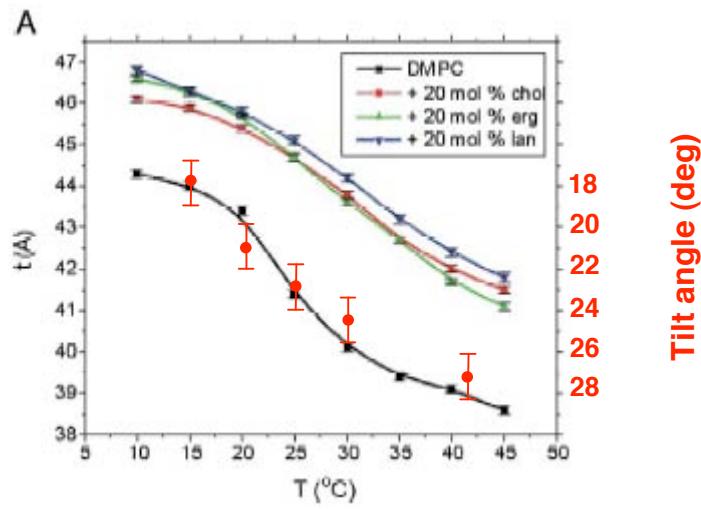
Mohamed N. Triba, Philippe F. Devaux, and Dror E. Warschawski

Unité Mixte de Recherche No. 7099, Centre National de la Recherche Scientifique, Institut de Biologie Physico-Chimique, Paris, France



# *TM helix tilt angle determined by bilayer thickness.*

## bilayer thickness vs. temperature



Temperature: Pencer et al 2005  
Addition of POPC: Triba et al 2006

# *Parallel alignment without lanthanides.*

Biphenyl Bicelle Disks Align Perpendicular to Magnetic Fields on Large Temperature Scales: A Study Combining Synthesis, Solid-State NMR, TEM, and SAXS

Cécile Loudet,\* Sabine Manet,\* Stéphane Gineste,<sup>†</sup> Reïko Oda,\* Marie-France Achard,<sup>†</sup> and Erick J. Dufourc\*

\*UMR 5248 CBMN, CNRS-Université Bordeaux 1-ENITAB, Institut Européen de Chimie et Biologie, Pessac, France; and

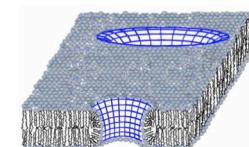
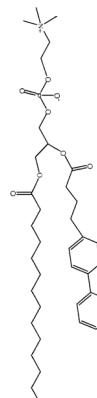
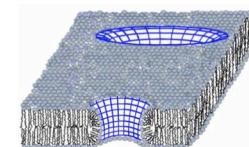
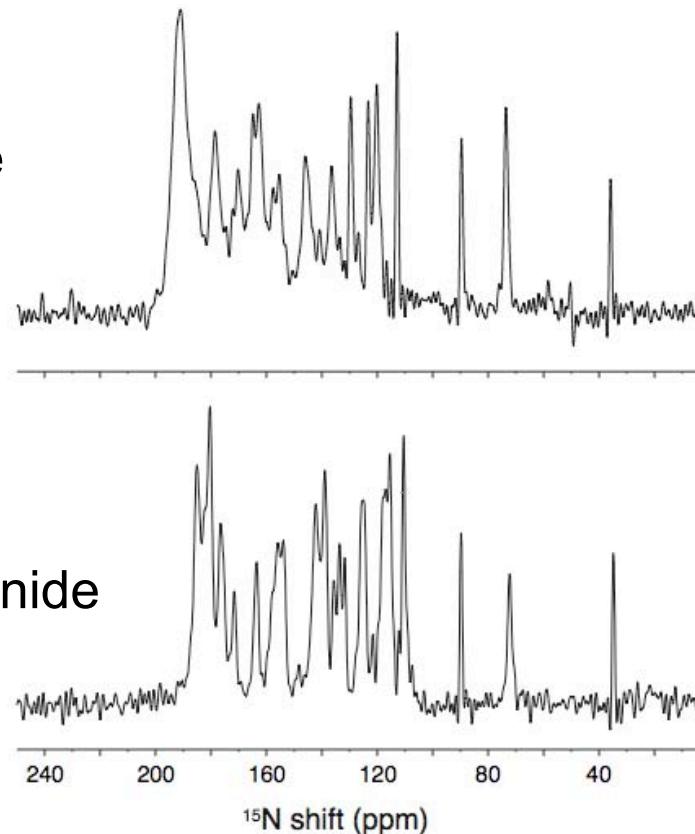
<sup>†</sup>UPR 8641, Centre de Recherche Paul Pascal, CNRS, Pessac, France

$q = 3.2$  bicelles

DMPC with lanthanide

TBBPC without lanthanide

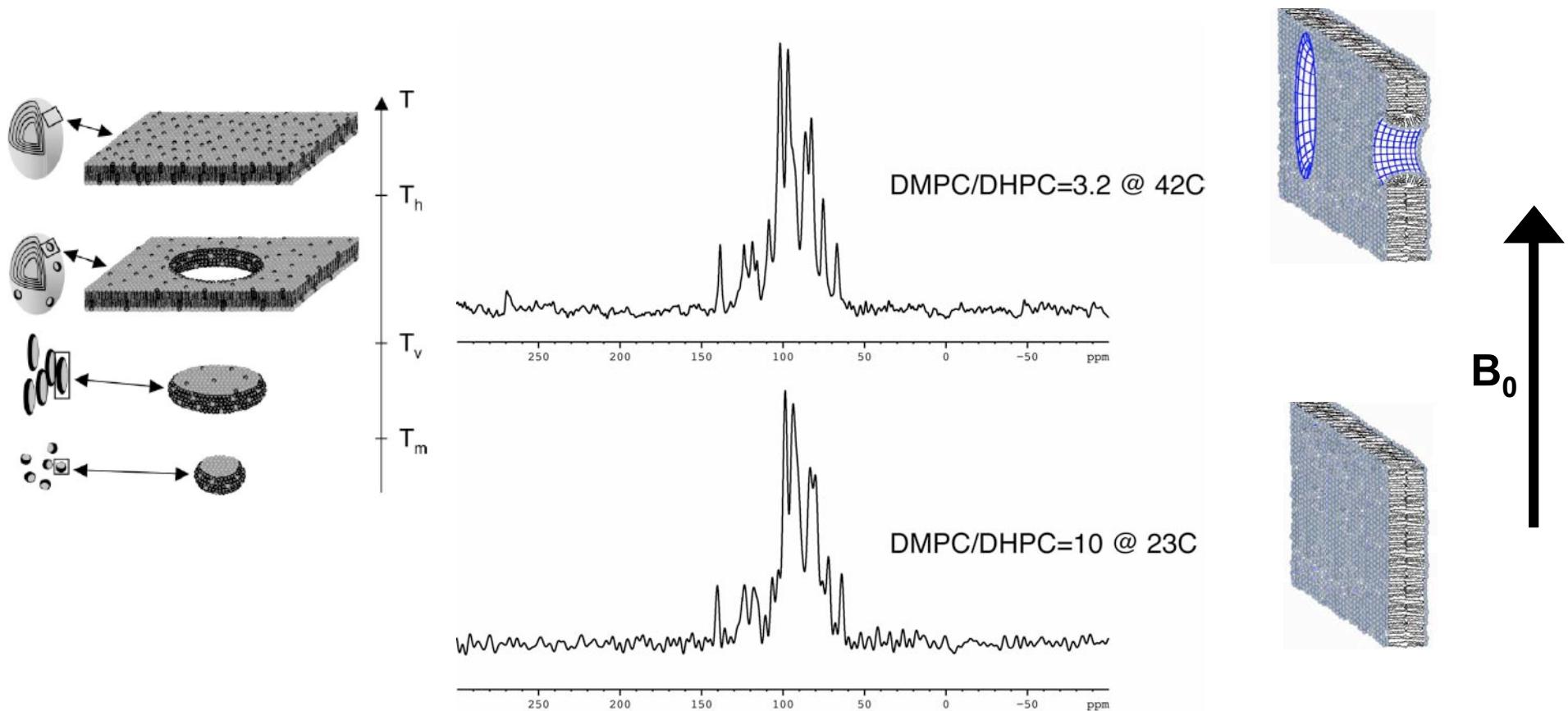
Biphenyl group confers positive magnetic anisotropy



# Reinvestigation by Phosphorus NMR of Lipid Distribution in Bicelles

Mohamed N. Triba, Dror E. Warschawski, and Philippe F. Devaux

Unité Mixte de Recherche No. 7099, Centre National de la Recherche Scientifique, Institut de Biologie Physico-Chimique, Paris, France



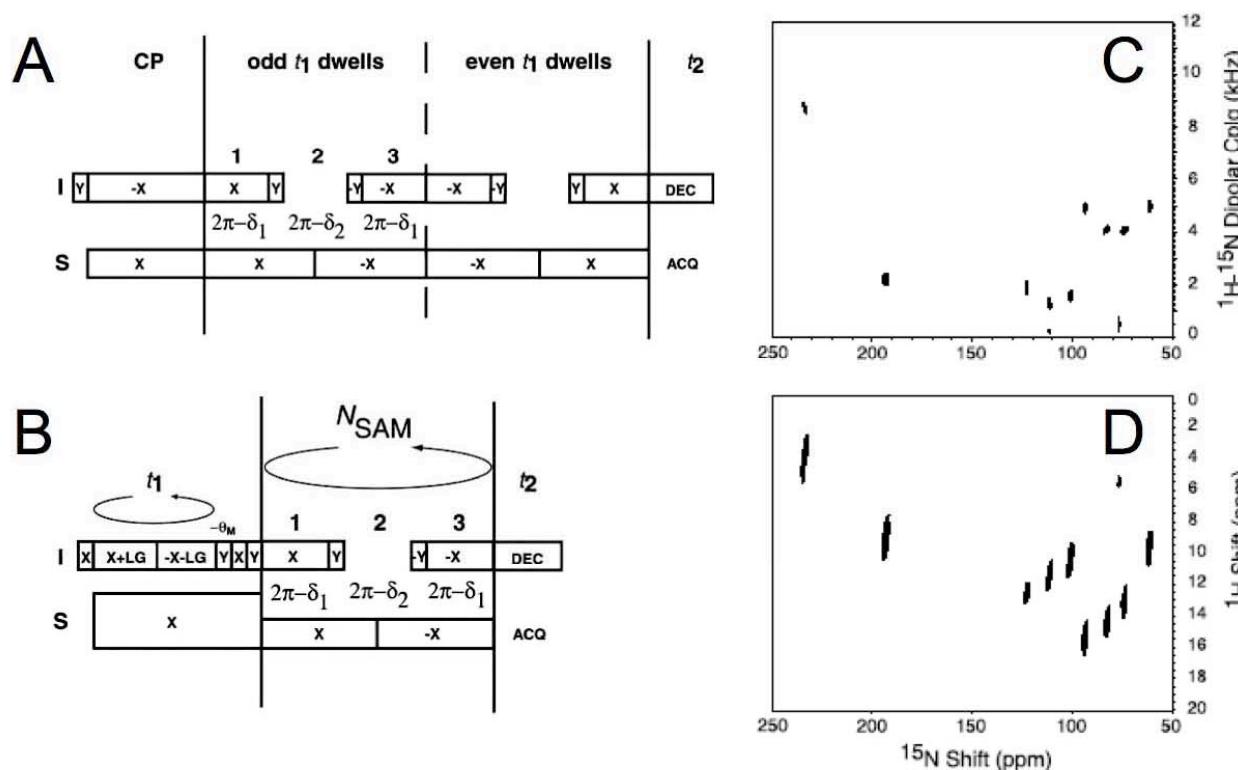
2005

*Double resonance experiments.*

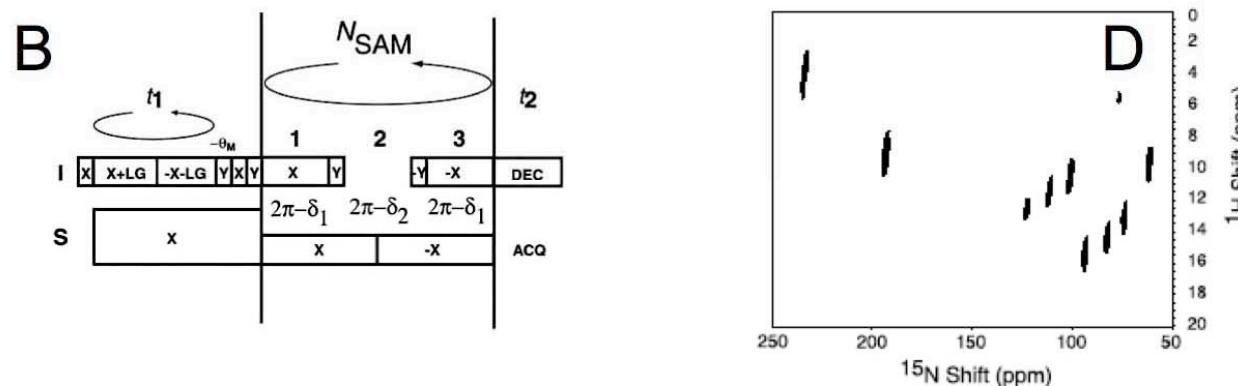
# Selective averaging for high-resolution solid-state NMR spectroscopy of aligned samples

Alexander A. Nevzorov, Stanley J. Opella \*

Department of Chemistry and Biochemistry, University of California, San Diego, 9500 Gilman Drive, La Jolla, CA 92093-0307, USA



SLF

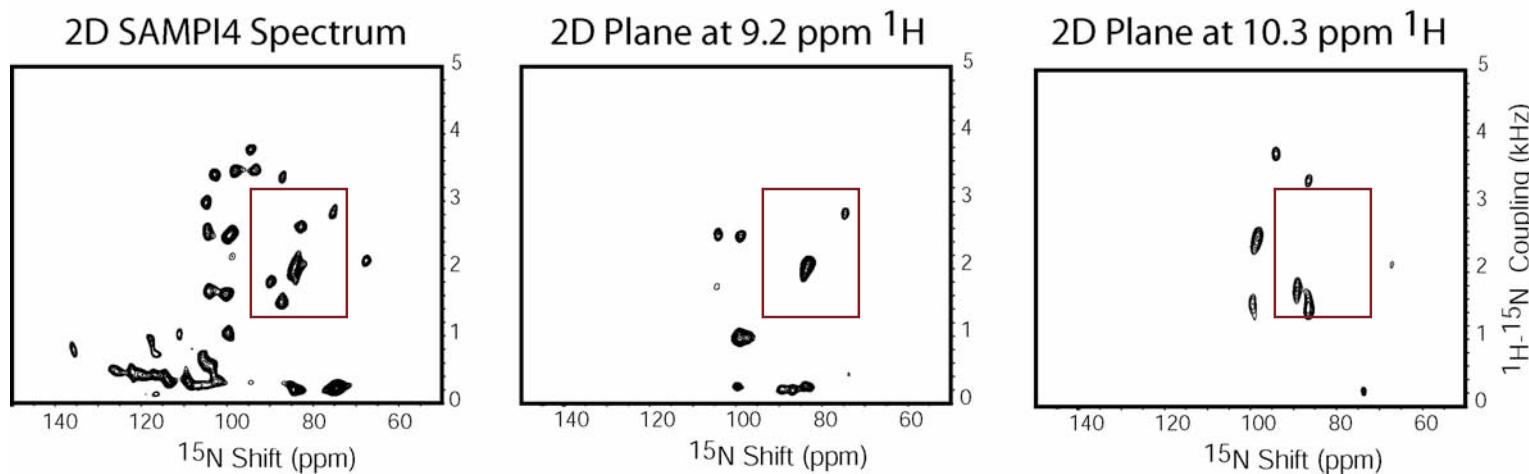
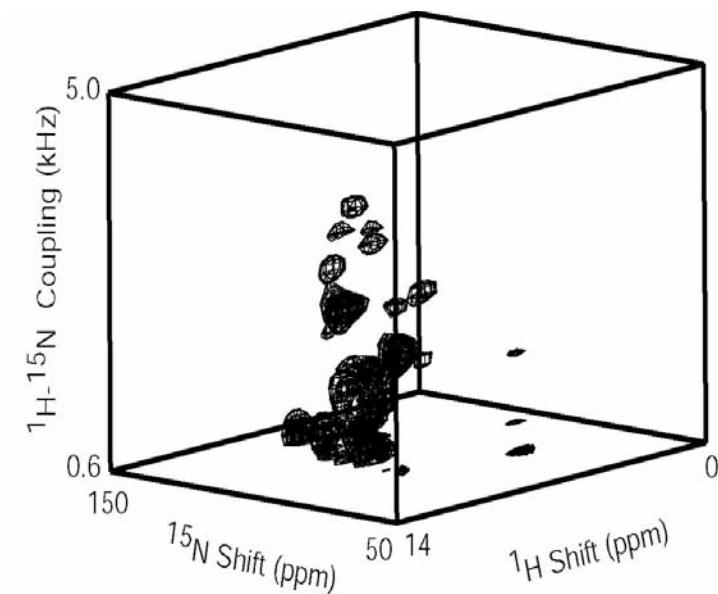
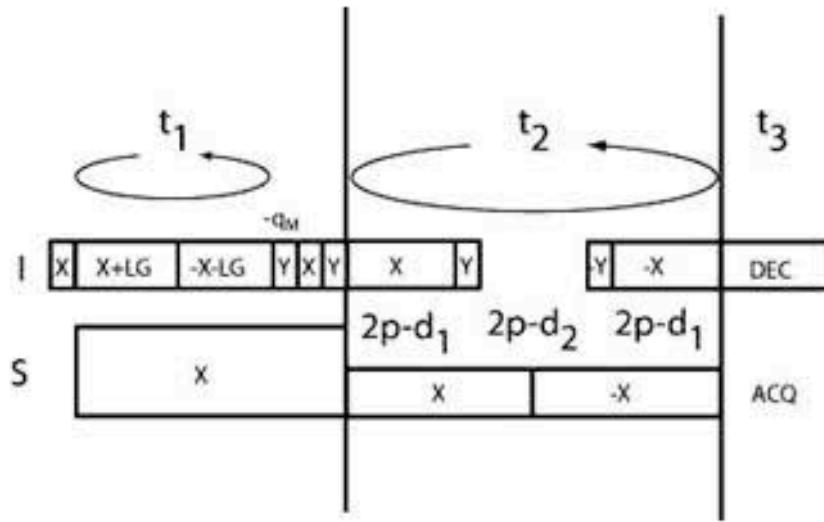


HETCOR

at 900 MHz  $^1\text{H}$  shift spans 14 kHz and  $^{15}\text{N}$  shift spans 16 kHz

2007

# SAMPI4 based $^1\text{H}$ Shift/ $^1\text{H}$ - $^{15}\text{N}$ Coupling/ $^{15}\text{N}$ Shift experiment at 900 MHz on a magnetically aligned bicelle sample.



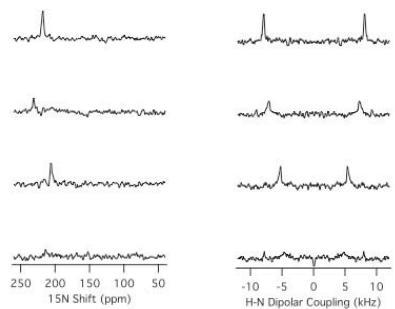
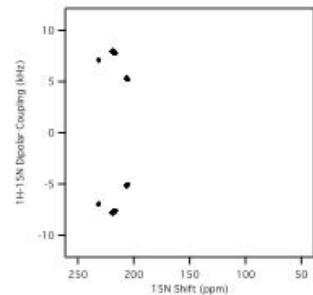
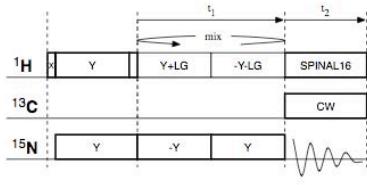
# Proton-detected separated local field spectroscopy

Chin H. Wu, Stanley J. Opella \*

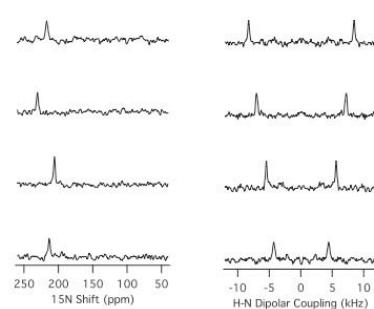
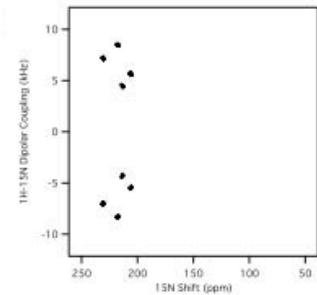
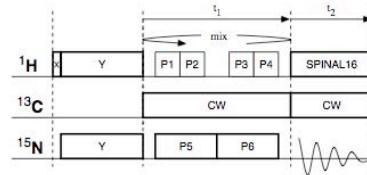
2008

Department of Chemistry and Biochemistry, 9500 Gilman Drive, University of California, San Diego, La Jolla, CA 92093-0307, United States

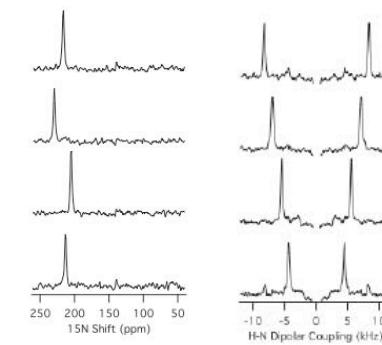
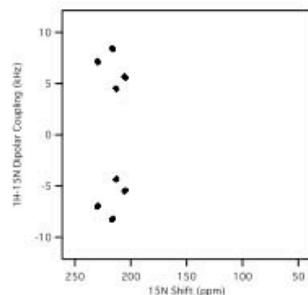
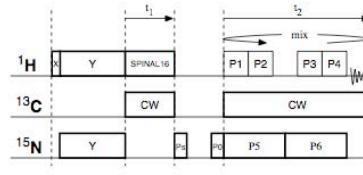
$^{15}\text{N}$ -detected PISEMA



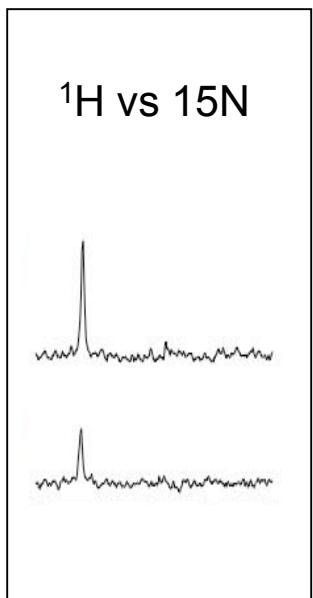
$^{15}\text{N}$ -detected PISEMO



$^1\text{H}$ -detected PISEMO



$^1\text{H}$  vs  $^{15}\text{N}$

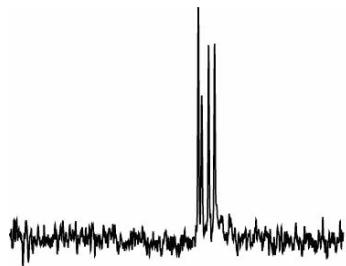


*Triple resonance experiments.*

## $^{13}\text{C}$ vs $^{15}\text{N}$ detection of $^{15}\text{N}$ , $^{13}\text{C}\alpha$ labeled peptides.

single crystal of N-acetyl-leucine

8 scans



**$^1\text{H} \rightarrow ^{13}\text{C}$  (detect)**

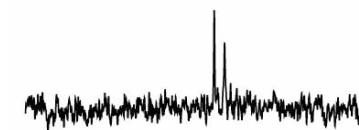
8 scans



**$^1\text{H} \rightarrow ^{15}\text{N}$  (detect)**

**$^1\text{H} \rightarrow ^{15}\text{N} \rightarrow ^{13}\text{C}$  (detect)**

8 scans

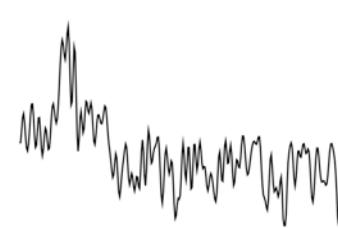
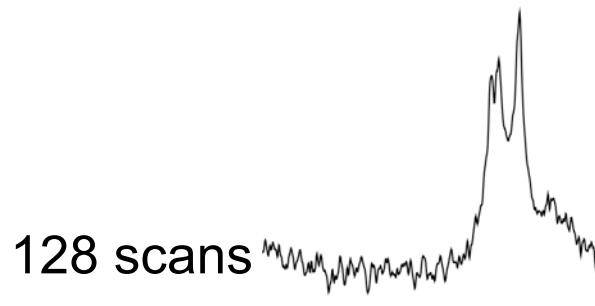


aligned sample of Leu (4 sites)  
labeled Pf1 bacteriophage

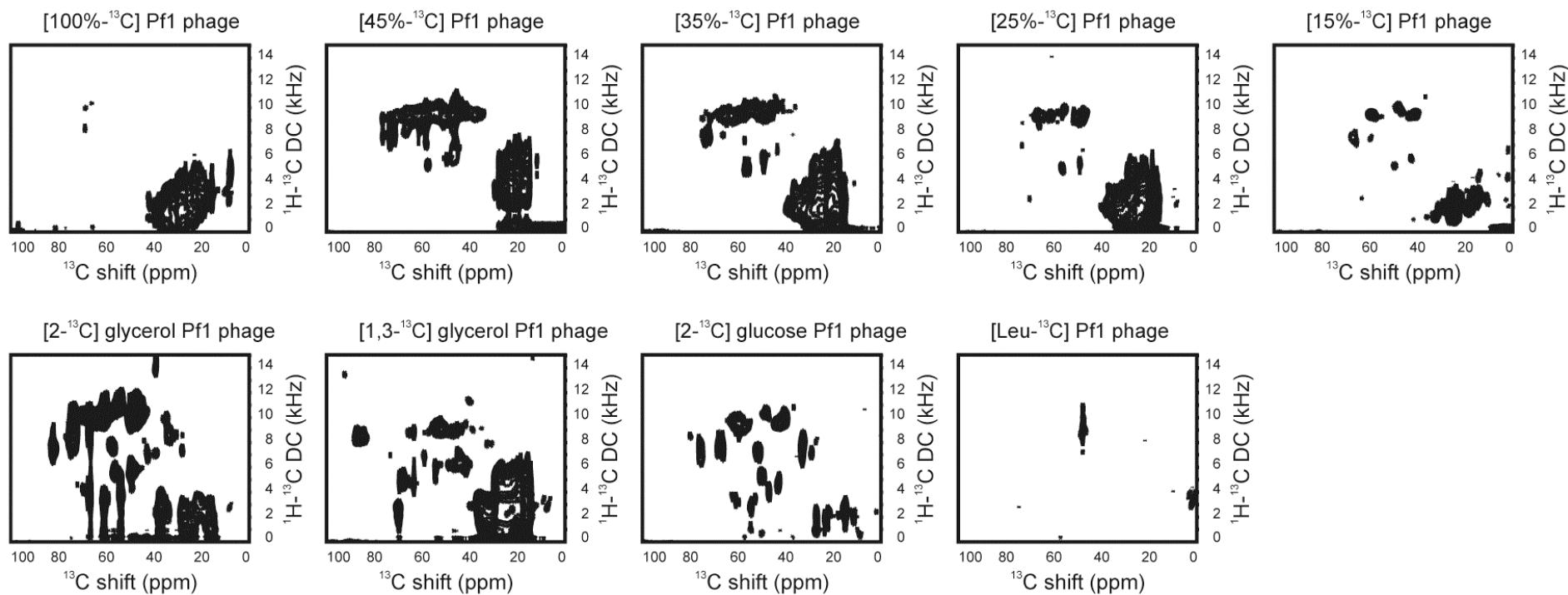
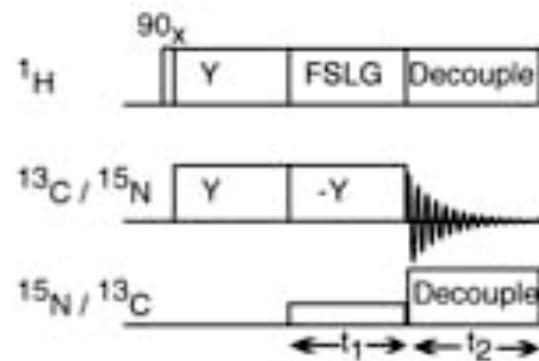
128 scans

128 scans

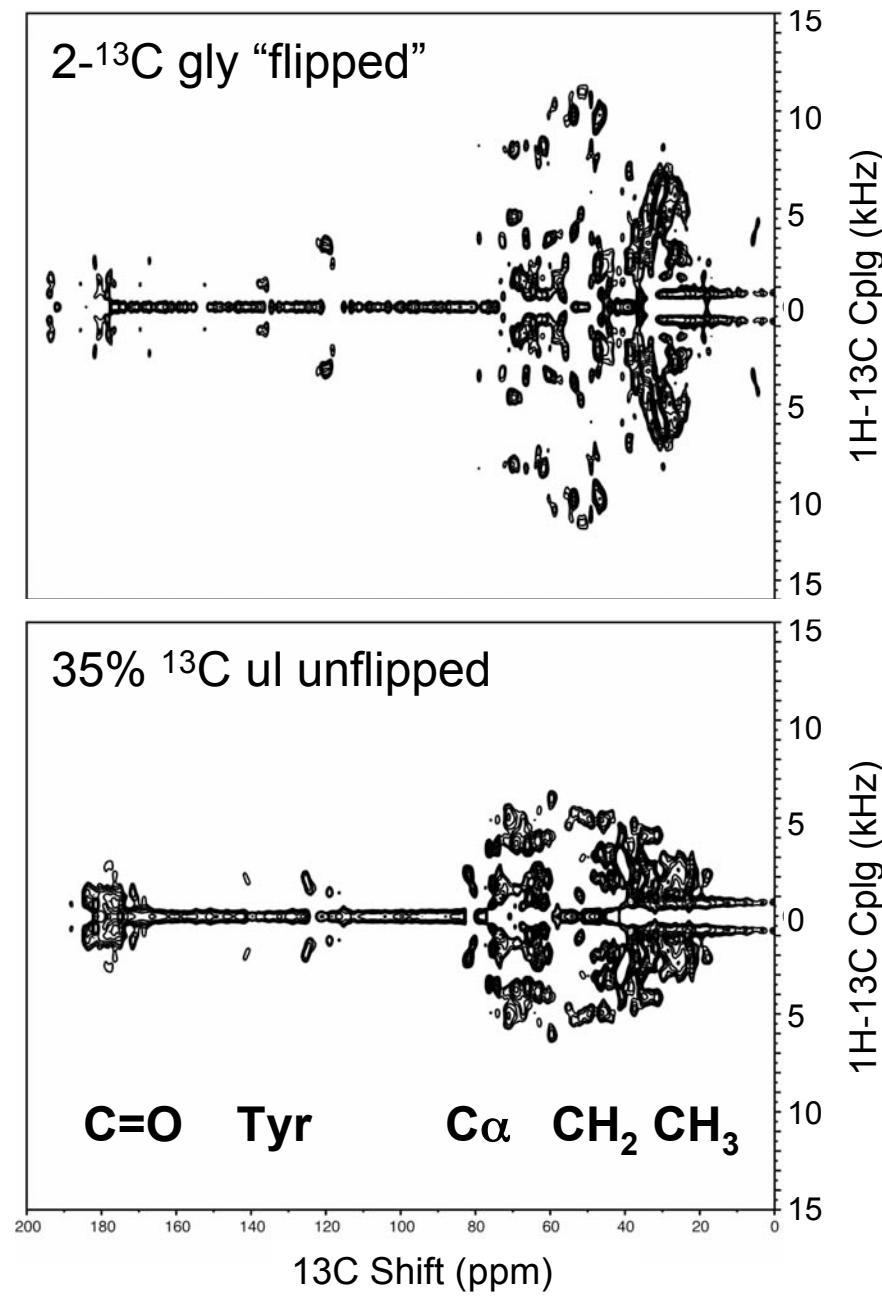
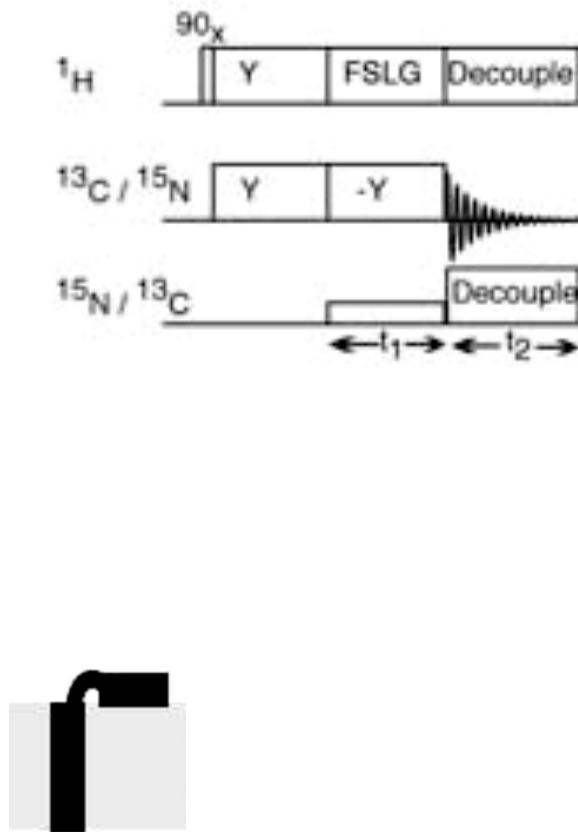
500 scans



# $^1\text{H}$ - $^{13}\text{C}$ PISEMA of $^{13}\text{C}$ and $^{15}\text{N}$ labeled Pf1 coat protein in aligned bacteriophage.

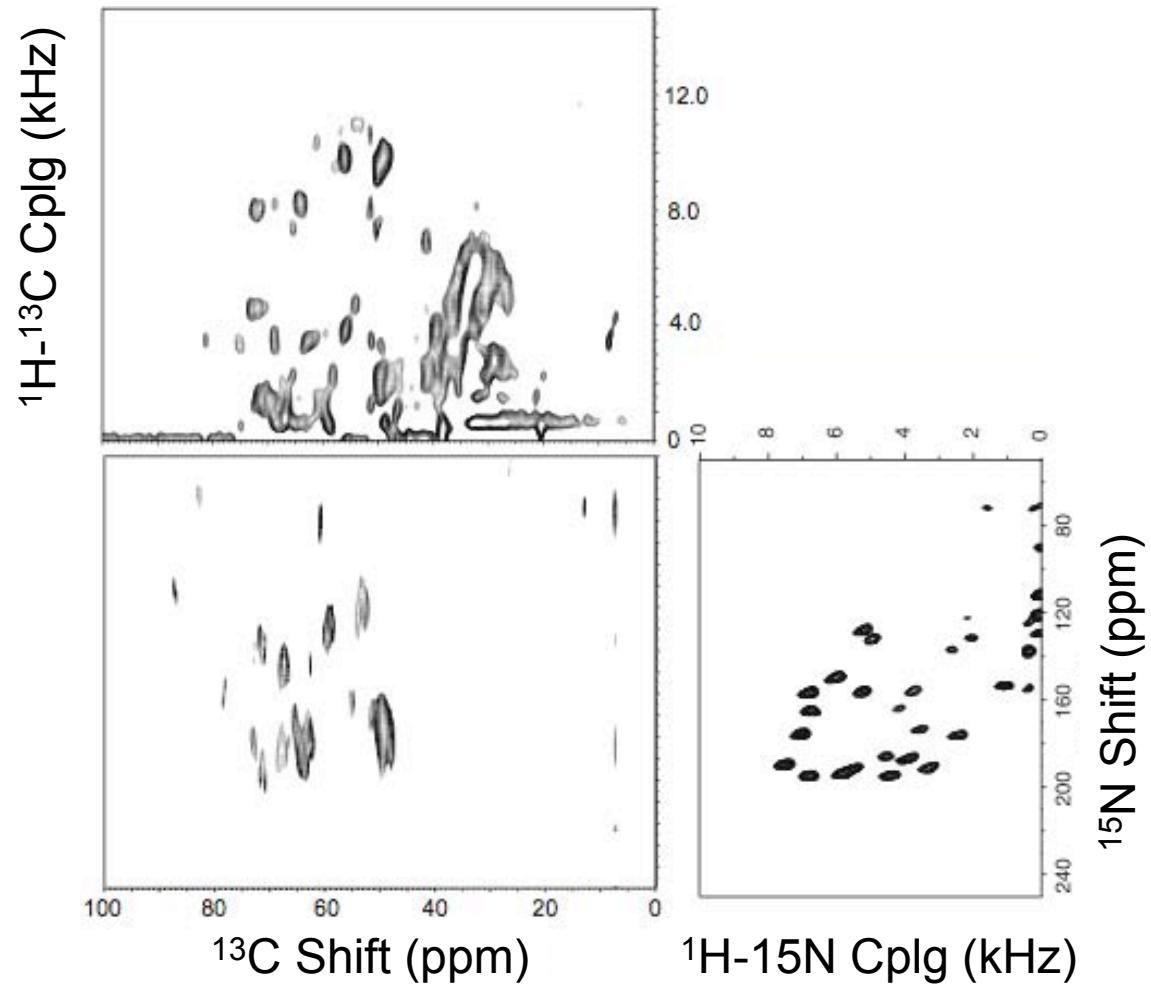
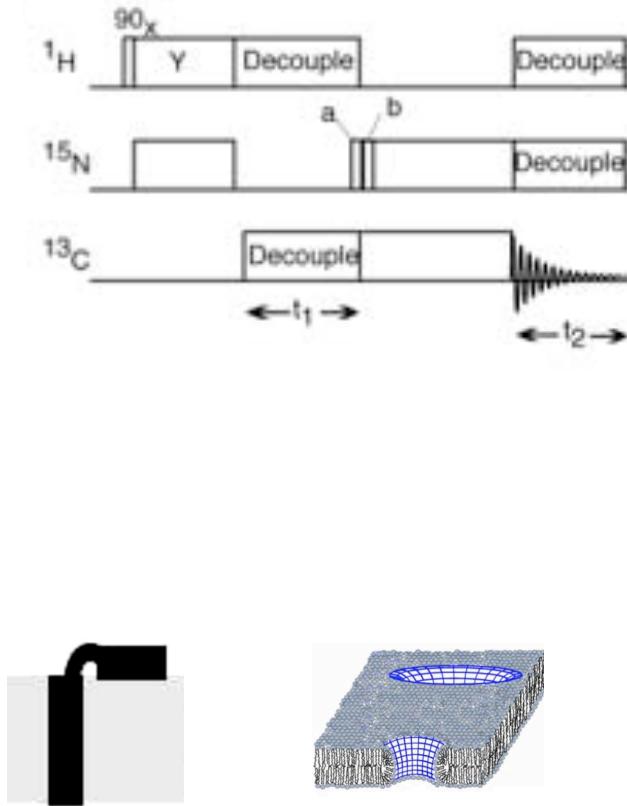


# $^1\text{H}$ - $^{13}\text{C}$ PISEMA of Pf1 coat protein in bicelles.



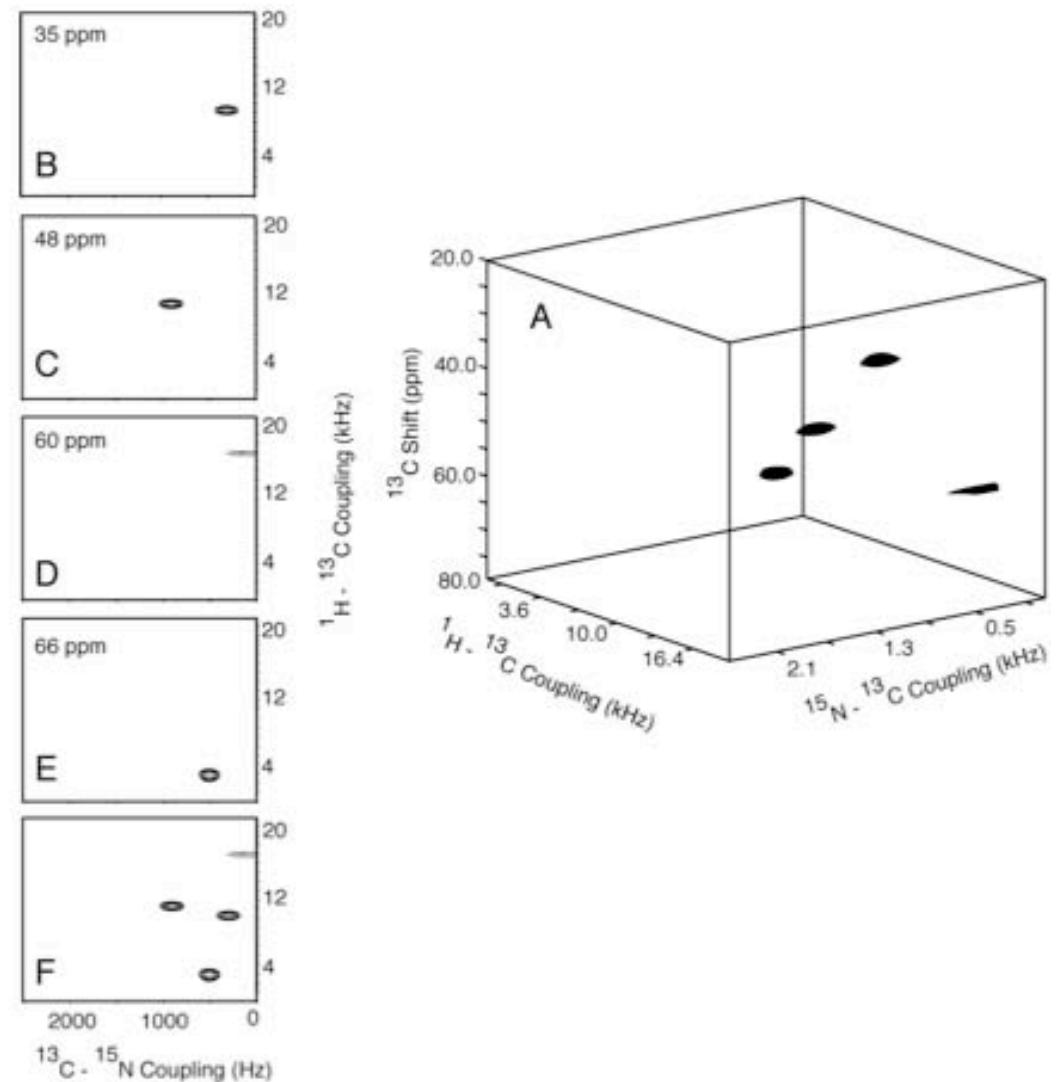
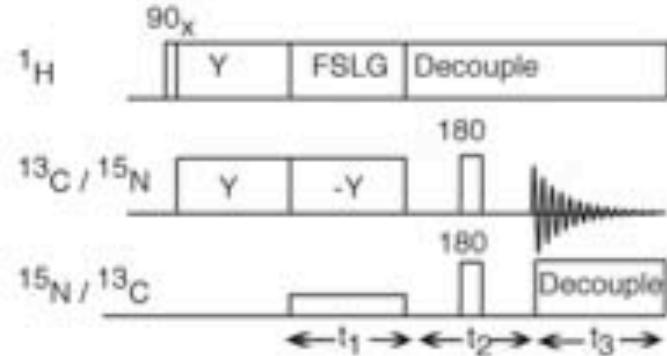
$^1\text{H}$ - $^{13}\text{C}$  PISEMA /  $^1\text{H}$ - $^{15}\text{N}$  PISEMA /  $^{13}\text{C}/^{15}\text{N}$  HETCOR.

2- $^{13}\text{C}$  glycerol/ul  $^{15}\text{N}$  labeled Pf1 coat protein in “flipped” bicelles.



# *Three-dimensional triple-resonance spectrum of a $^{13}\text{C}\alpha$ , $^{15}\text{N}$ labeled model peptide crystal.*

$^{13}\text{C}$  shift separated  $^1\text{H}$ - $^{13}\text{C}$ / $^{13}\text{C}$ - $^{15}\text{N}$  dipolar spectra

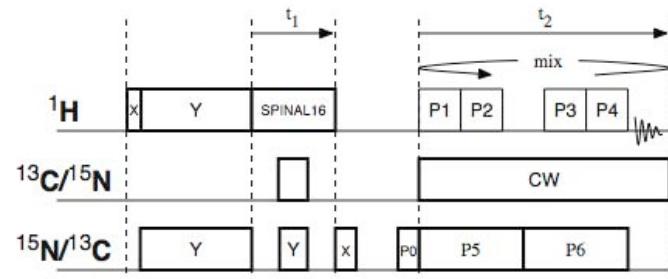


# Shiftless nuclear magnetic resonance spectroscopy

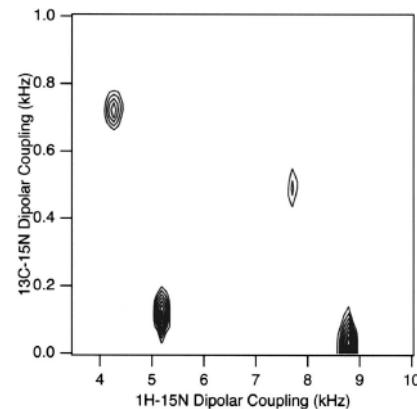
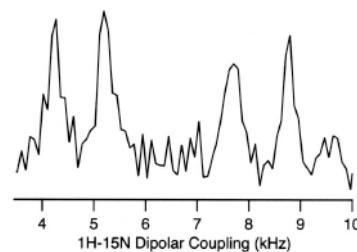
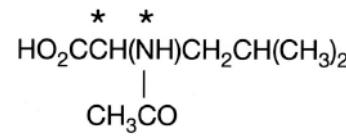
Chin H. Wu and Stanley J. Opella<sup>a)</sup>

Department of Chemistry and Biochemistry, University of California, San Diego, 9500 Gilman Drive,  
La Jolla, California 92093-0307, USA

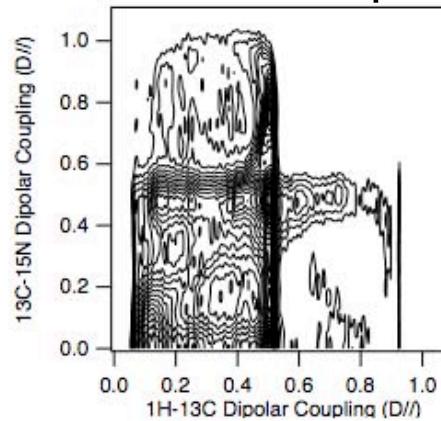
2008



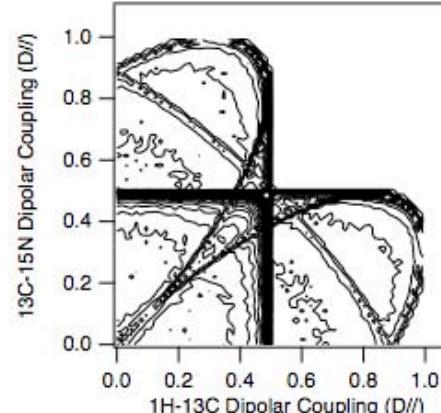
single crystal



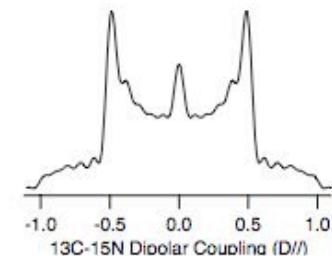
A



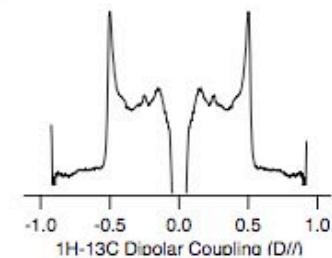
B



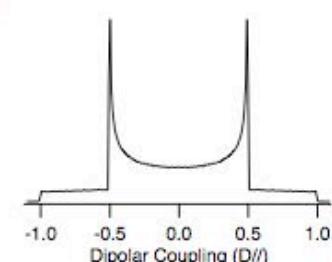
powder C



D



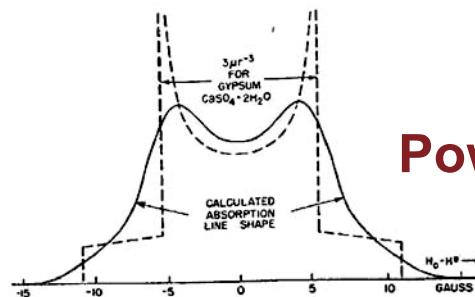
E



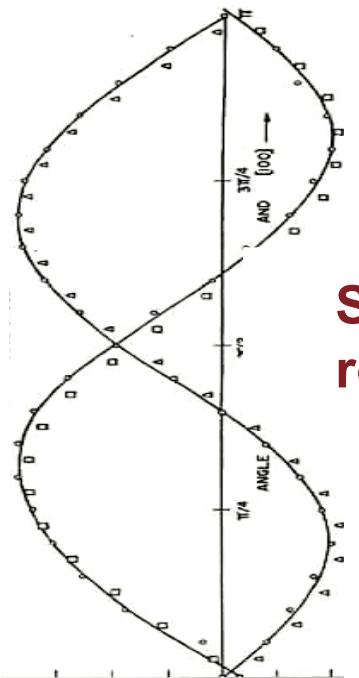
*The dipole-dipole interaction is anisotropic.*

Nuclear Resonance Absorption in Hydrated Crystals:  
Fine Structure of the Proton Line

G. E. PAKE\*



Powder pattern.



Single crystal  
rotation pattern.

1948

***Part II: Phospholipid bilayers are essential for biological relevance.***

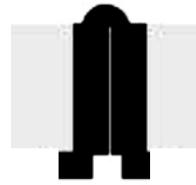
*Mercury transport membrane proteins with 2, 3, and 4 trans-membrane helices.*

**MerE**    **MerF**    **MerT**    **MerC**  
**2TM**    **2TM**    **3TM**    **4TM**

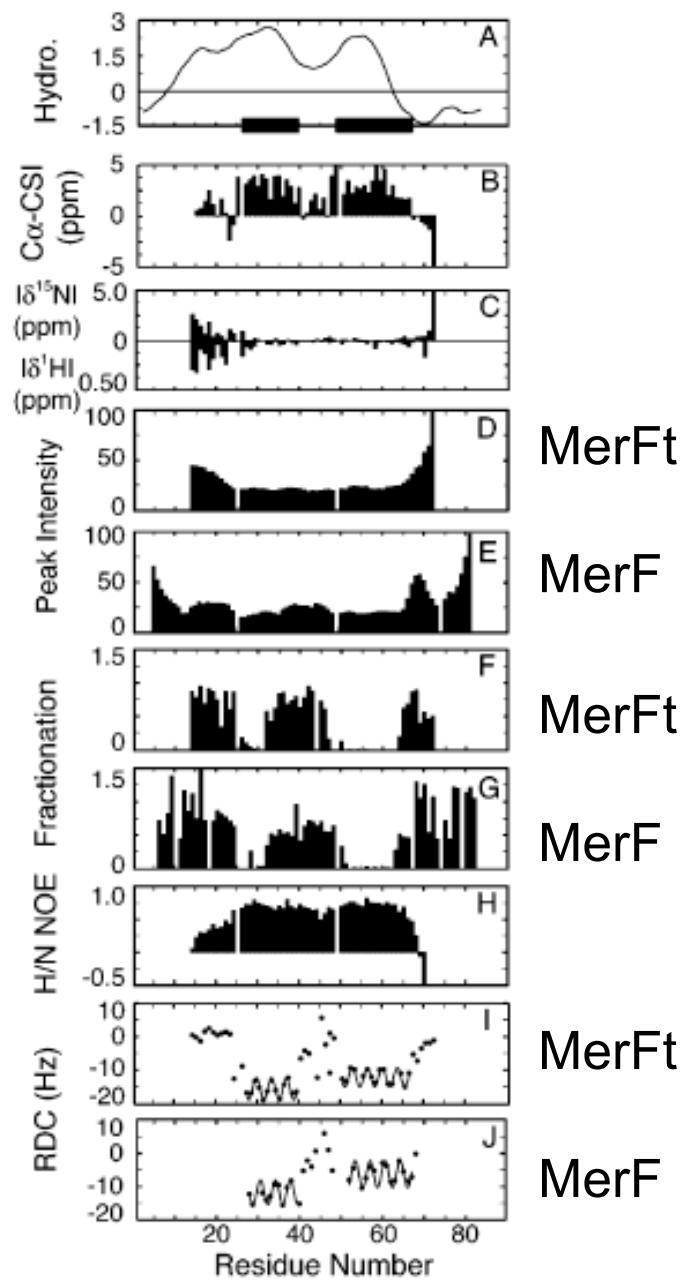
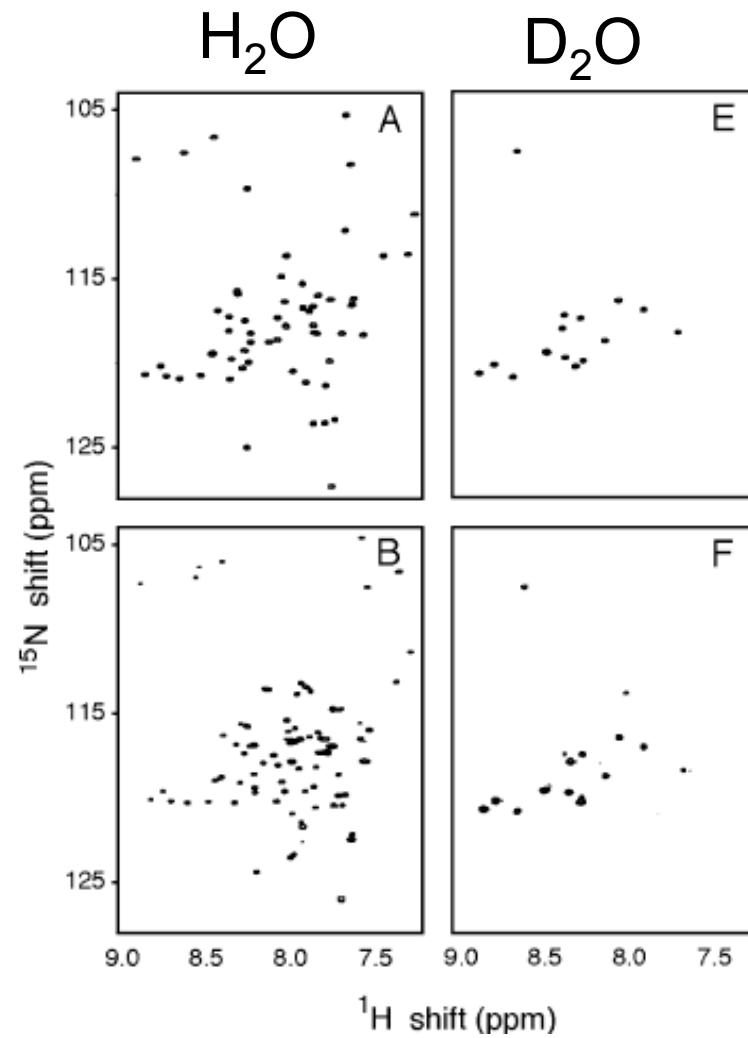
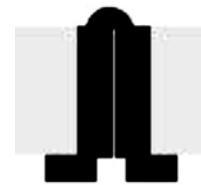


# *Solution NMR of the mercury transport membrane protein MerF in micelles.*

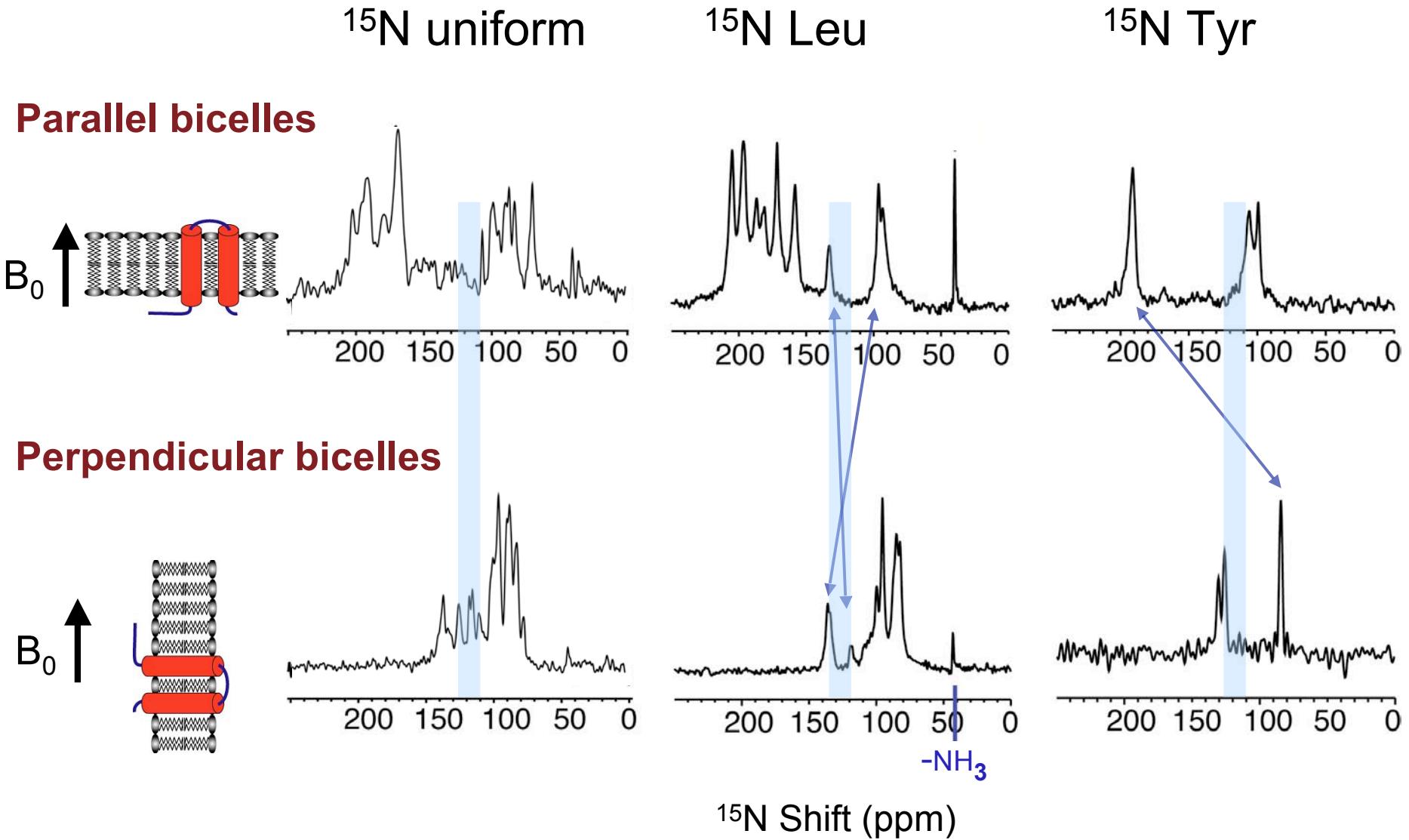
MerFt  
60 residues  
(truncated)



MerF  
80 residues  
(full length)



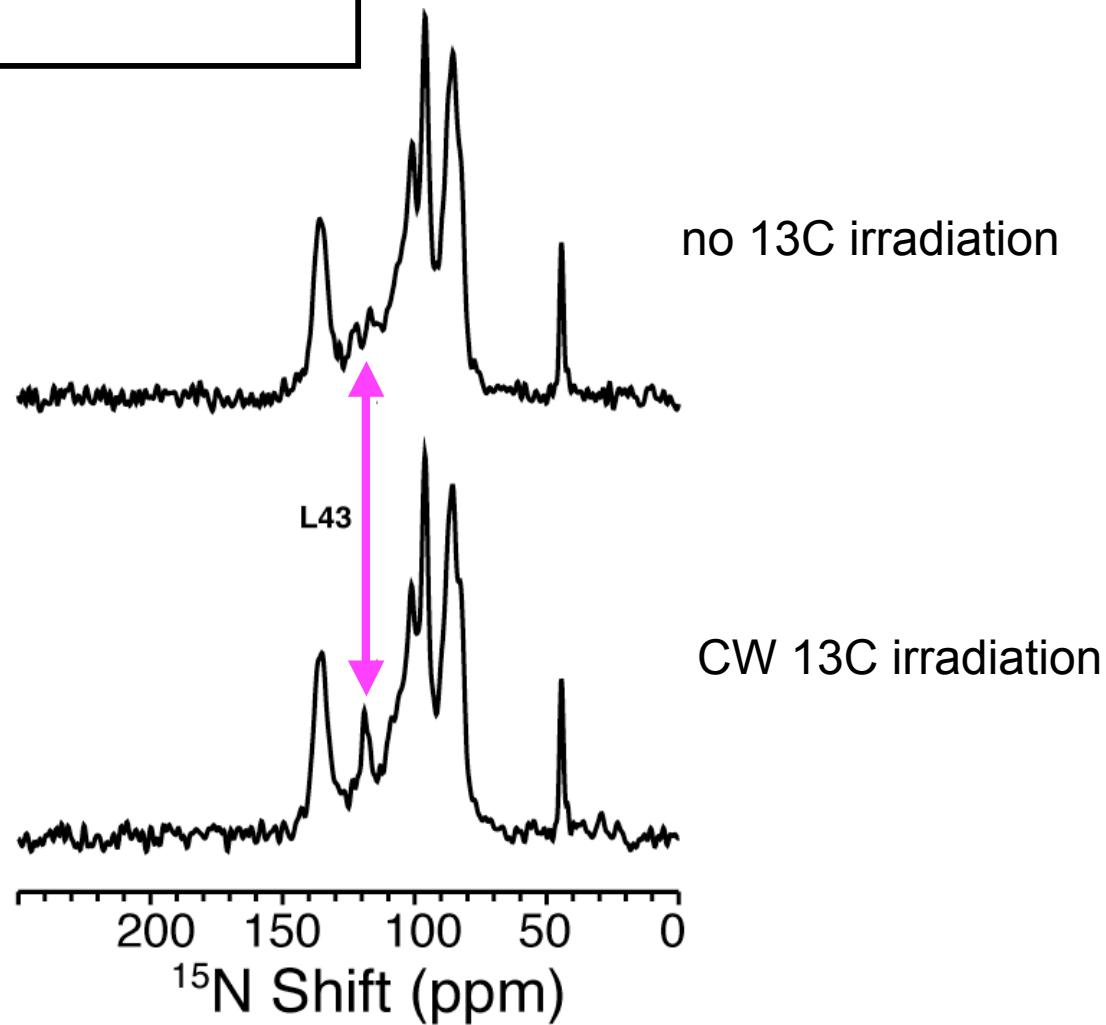
## *Effect of “flipping” on MerFt (60aa, 2TM) spectra.*



*<sup>13</sup>C decoupling of a selectively <sup>15</sup>N Leu (13 sites) and <sup>13</sup>C' Tyr (3 sites) labeled membrane protein.*

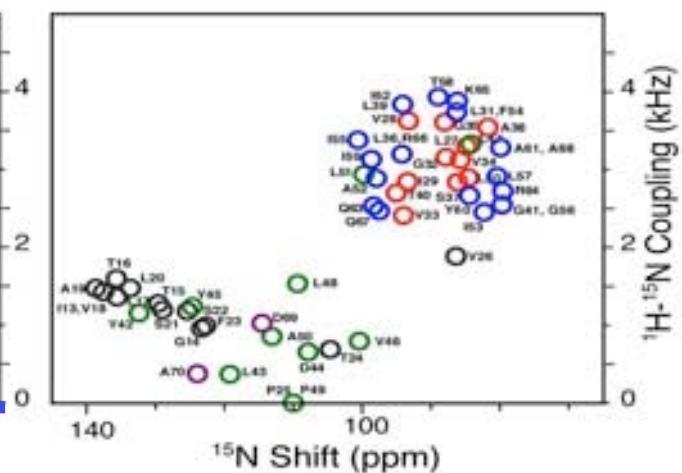
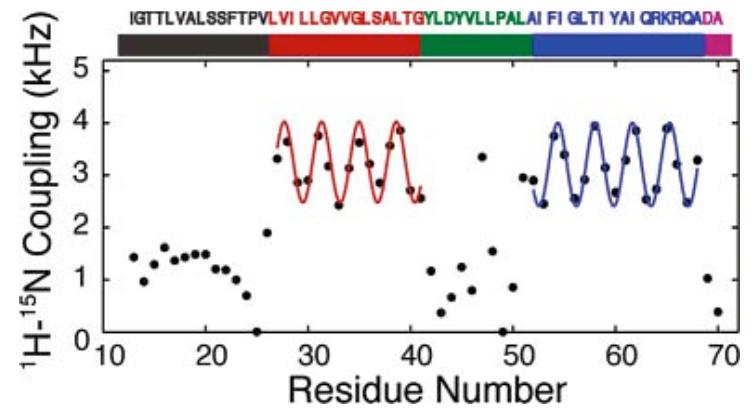
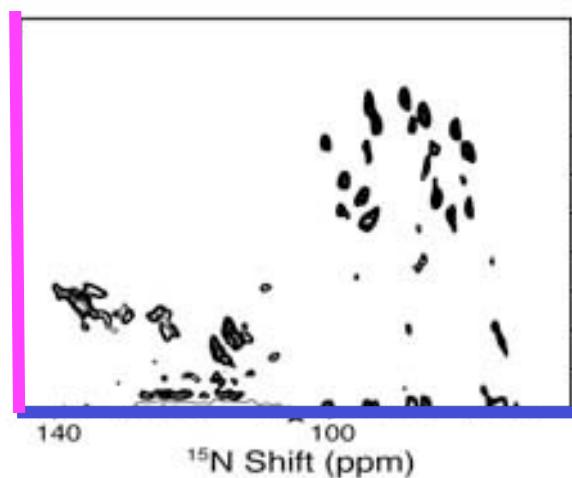
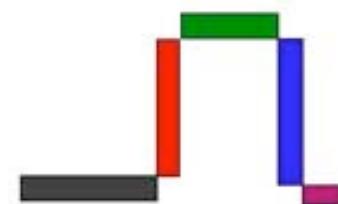
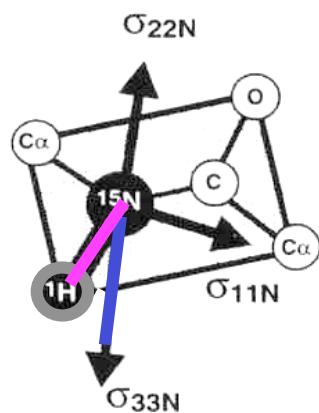
only one <sup>13</sup>C-<sup>15</sup>N bond in the protein

Tyr 42 - Leu 43

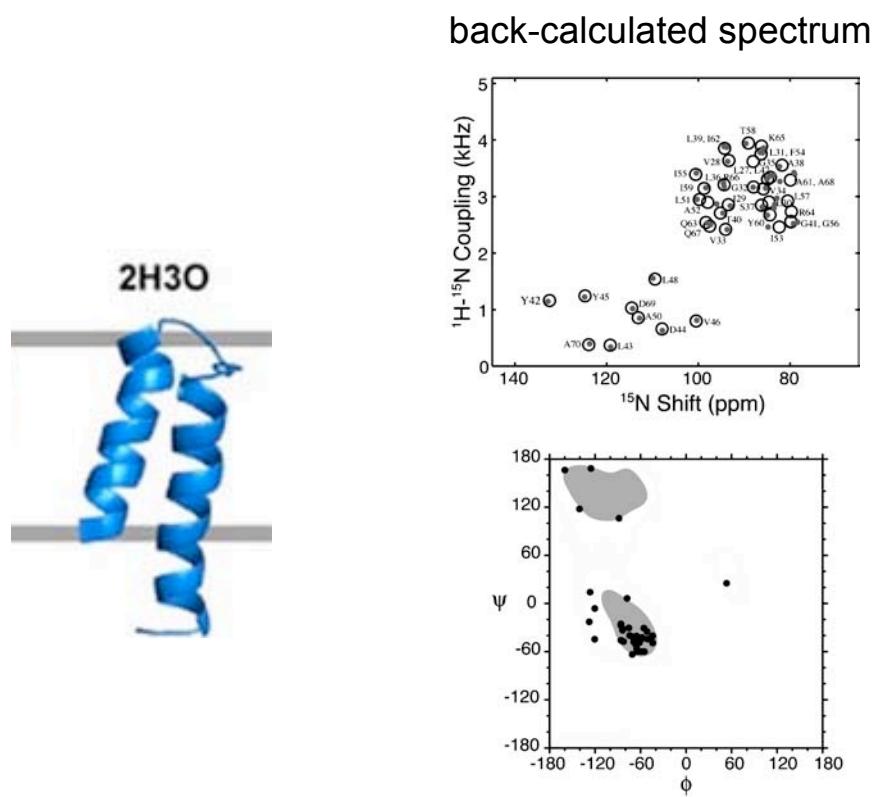
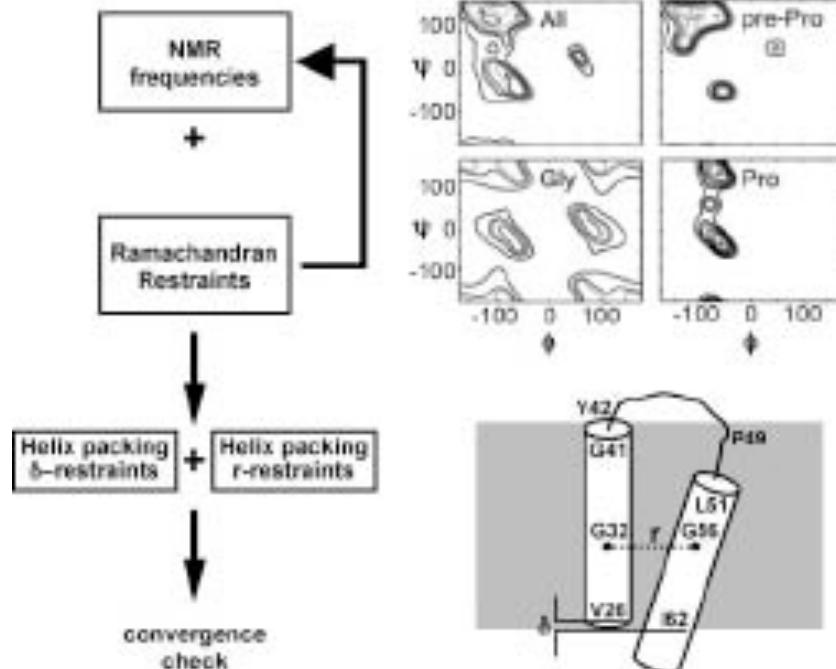


# *Resolution, measurement, and assignment of resonances.*

**MerFt (60 aa)**



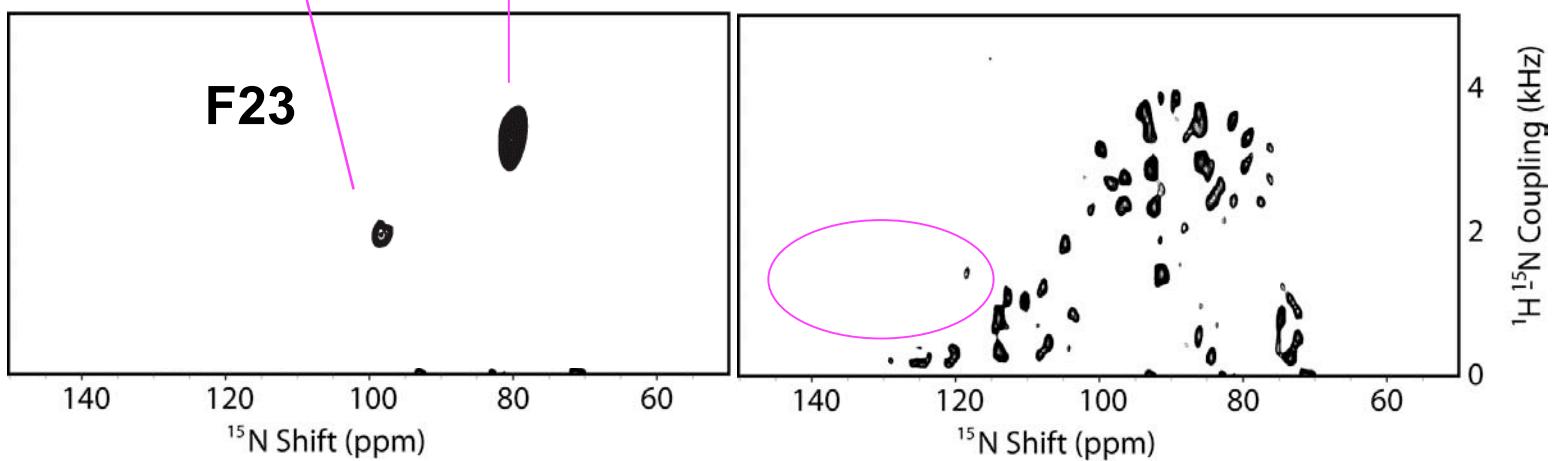
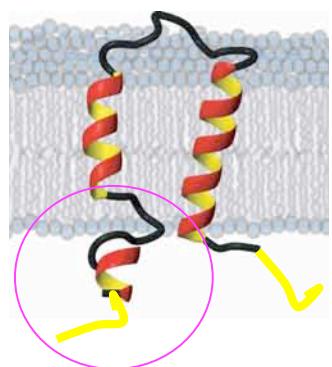
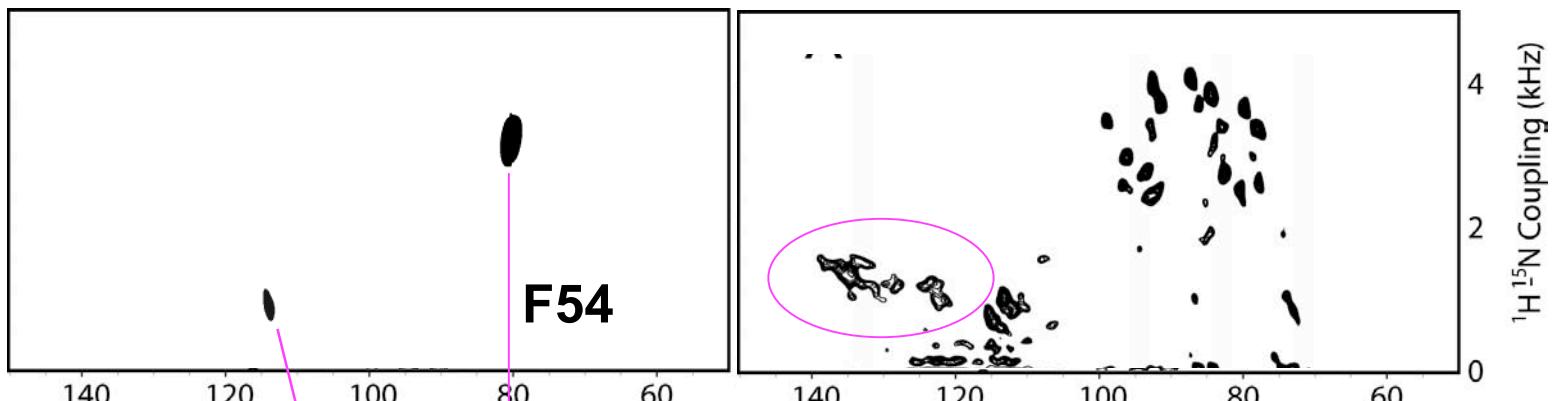
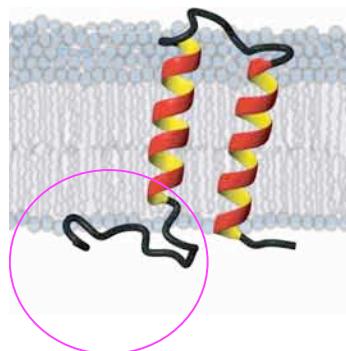
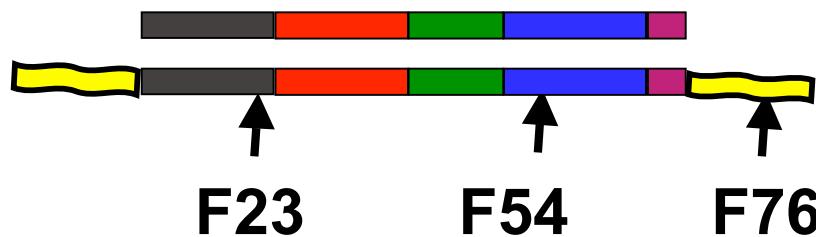
*Two orientationally dependent frequencies for each residue enable calculation of three-dimensional protein structures.*



# *Structure and dynamics of terminal domains of MerF.*

**MerFt (60 aa)**

**MerFm (80aa)**



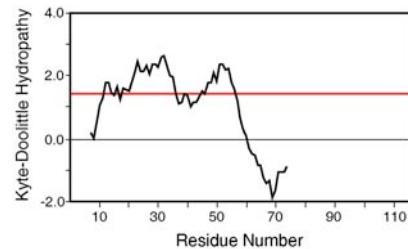
# *MerF* (80 aa, 2 TM) vs. chimeric *MerTf* (114 aa, 3 TM).

MerFm KDPKTLLRVSIIIGTTLVALSSFTPVLVILLGVVGLSALTGYLDYVLLPALAIFIGLTIYAIQRKROADASS---TPKFNGVKKZ-----  
 MerTf KDPKTLLRVSIIIGTTLVALSSFTPVLVILLGVVGLSALTGYLDYVLLPALAIFIGLTIYAIQRKROADASSAASKPGEVSAIPQVRATYKLIFWGVAVLVLVALGFPYVPFFYZ

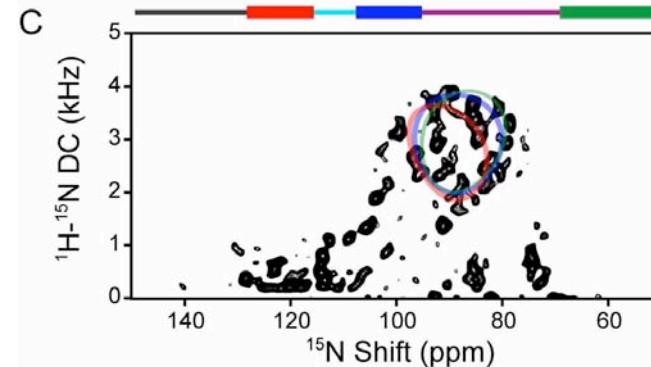
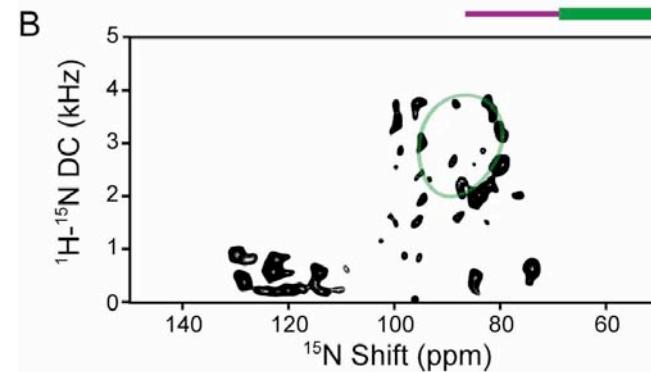
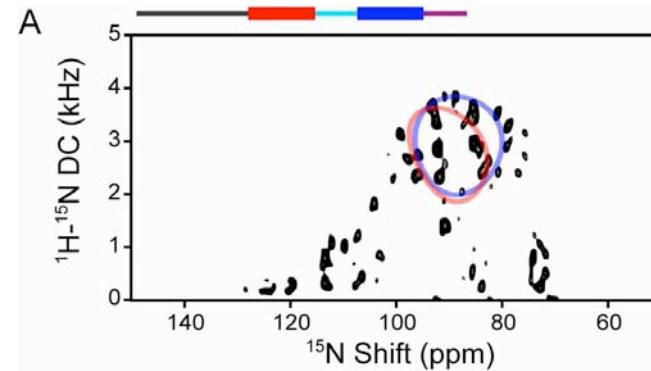
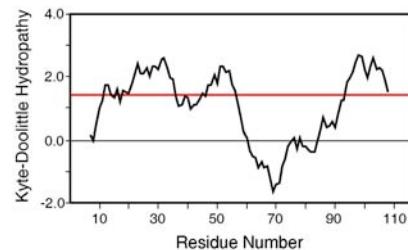
hydropathy plots



MerF

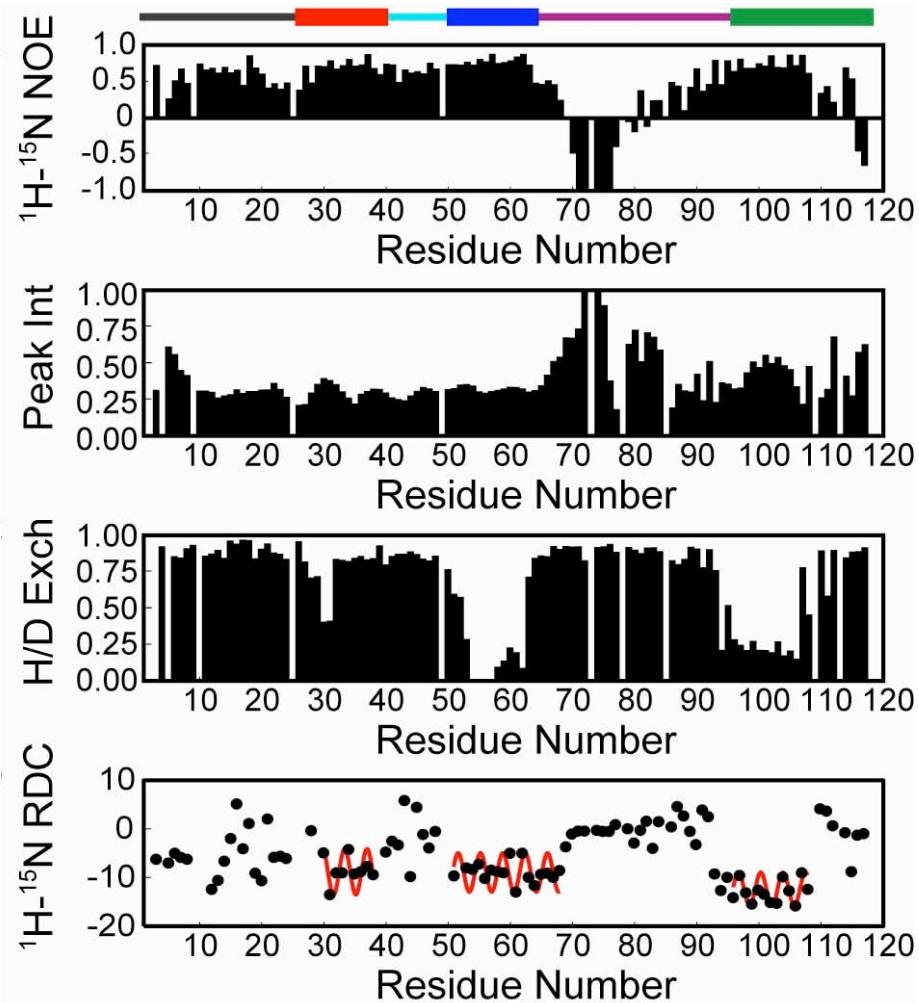


MerT

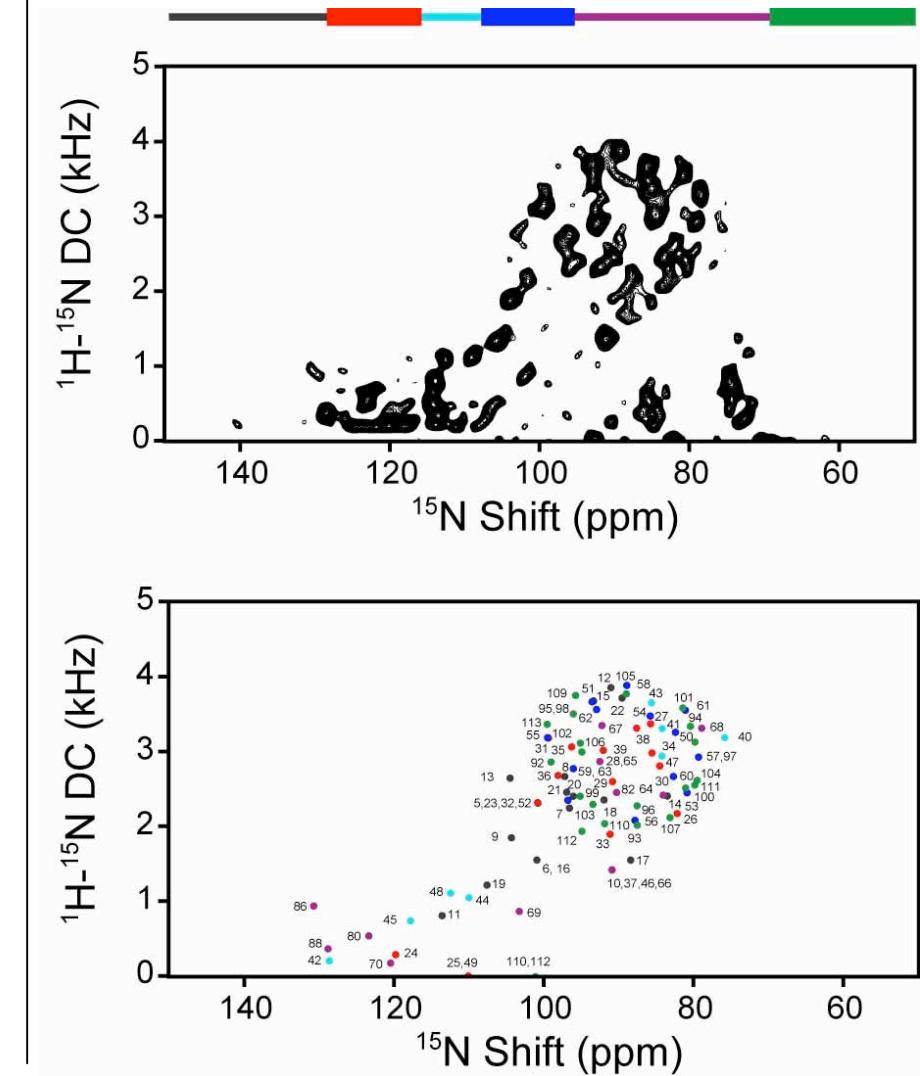


# *MerTf chimera in micelles vs. q=3.2 bicelles.*

## solution NMR

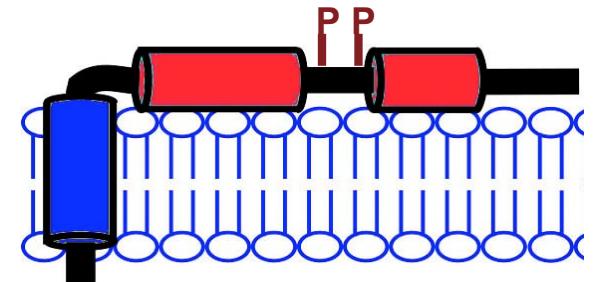


## solid-state NMR



## *Vpu of HIV-1.*

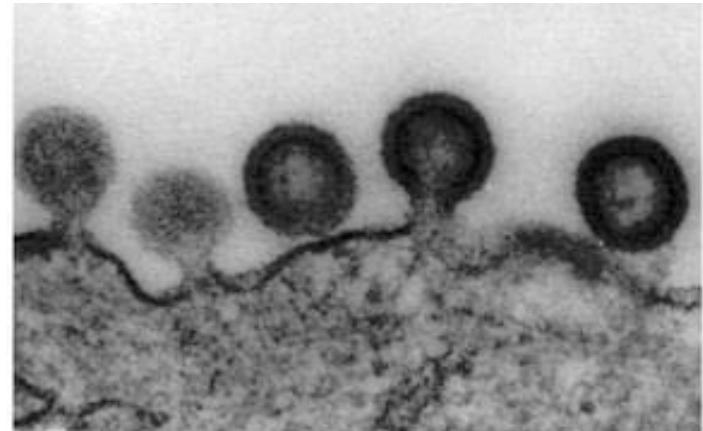
- 81 residue membrane protein.
- Enhances virus particle release from infected cells.
  - Associated with the trans-membrane domain.
  - Associated with ion channel activities.
  - Affected by “channel-blocking” drugs.
- Facilitates the degradation of CD4/gp160 complex.
  - Associated with the cytoplasmic domain.
  - Affected by phosphorylation of two conserved serines.



### **Vpu oligomer forms ion channels**

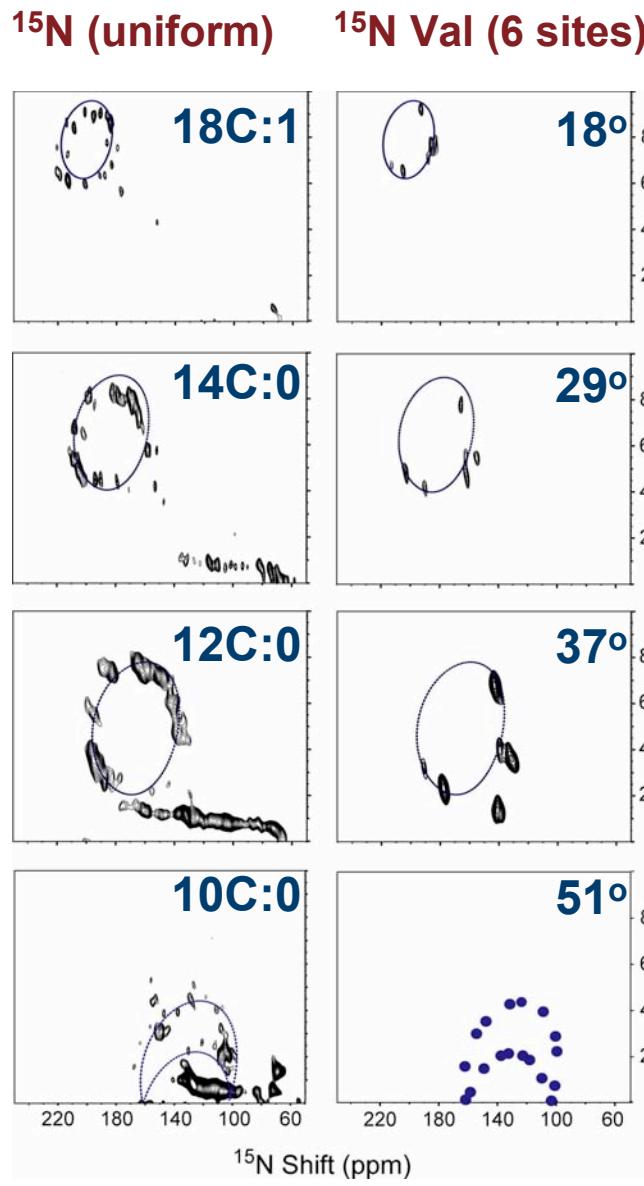


### **HIV budding from membrane**



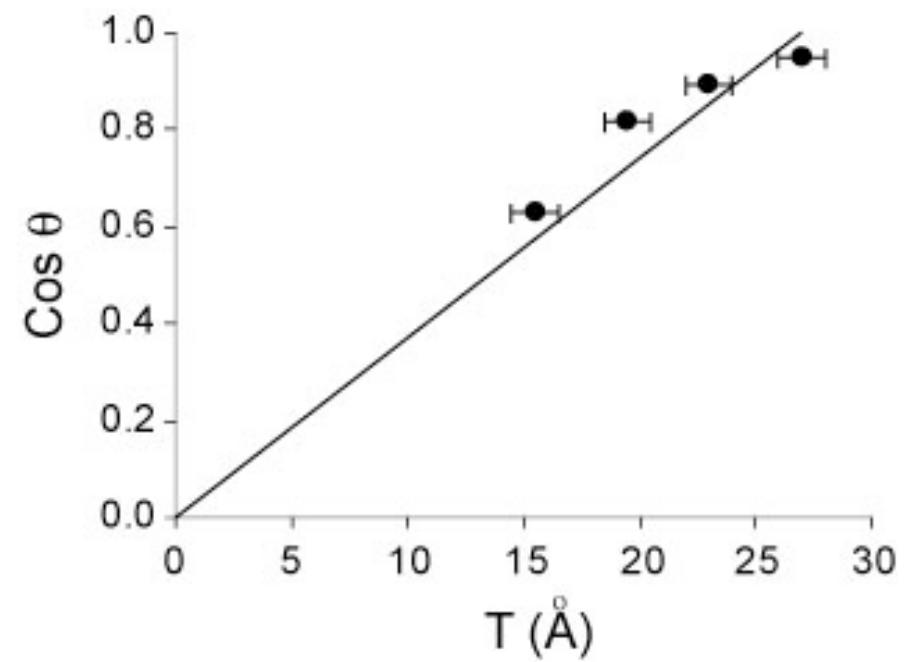
Ewart et al, Eur Biophys J 31, 26, 2002

# *Tilt angle compensates for hydrophobic mismatch. No change in rotation angle.*

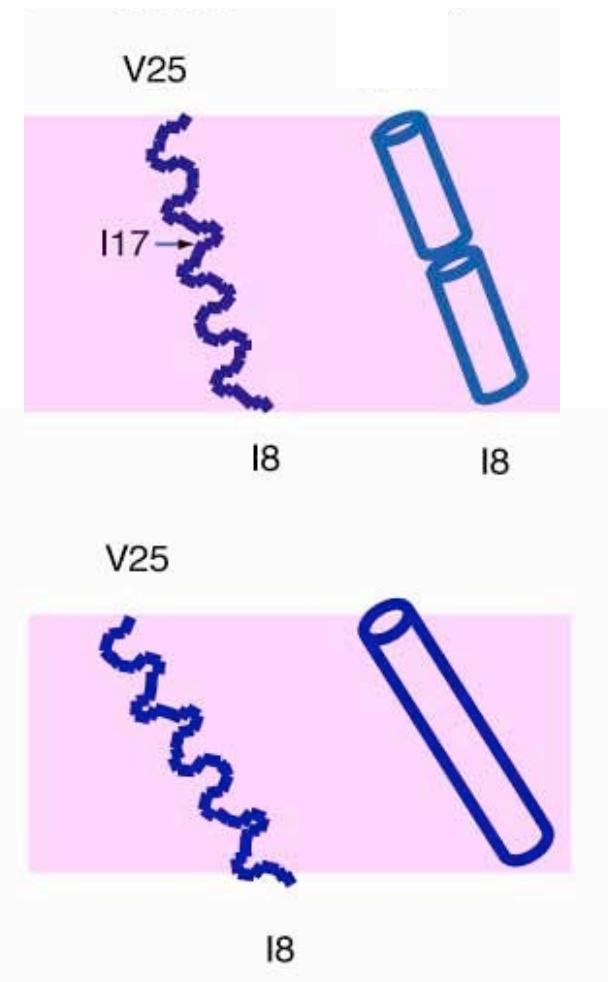
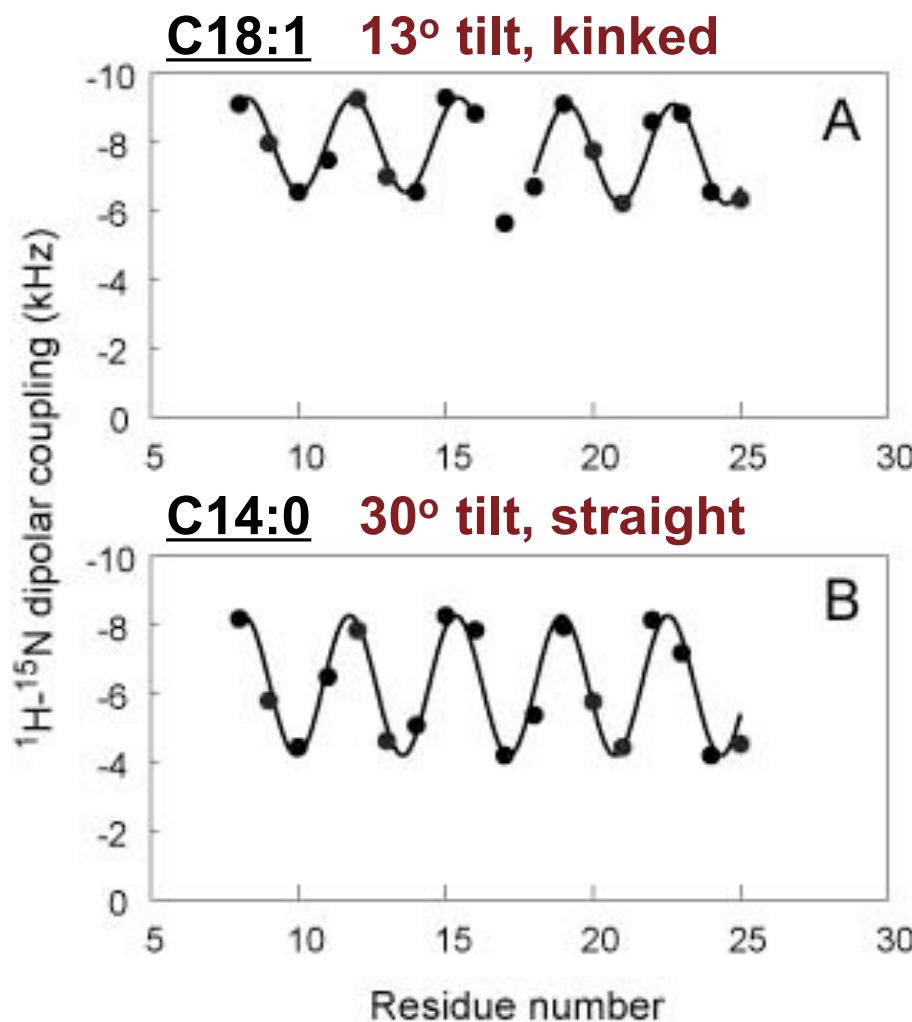


$$\Theta = \cos^{-1} \left( \frac{L}{T} \right)$$

A diagram showing a right-angled triangle with a horizontal base labeled 'L' and a vertical height labeled 'T'. The hypotenuse is labeled with the Greek letter Θ. A curved arrow indicates the angle Θ is at the vertex where the vertical side T meets the hypotenuse.



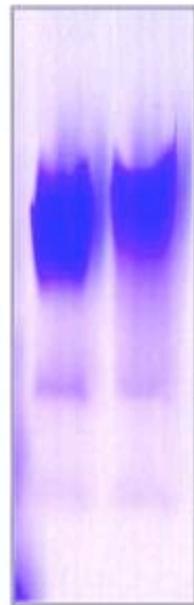
## *Lipids affect the structure of wild-type Vpu TM.*



*Gel electrophoresis suggests that Vpu forms a tetramer.*

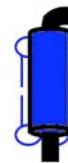
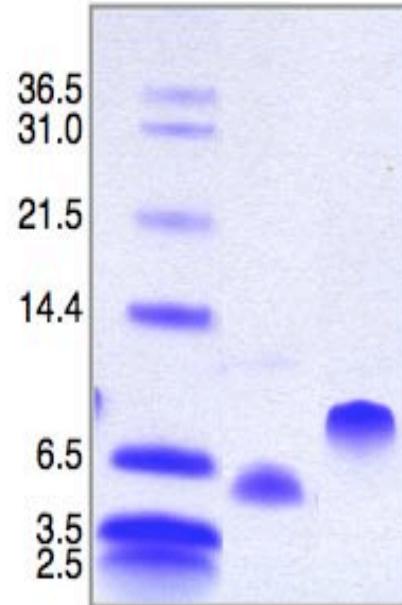
PFO PAGE

wild-type Vpu TM  
A18H Vpu TM

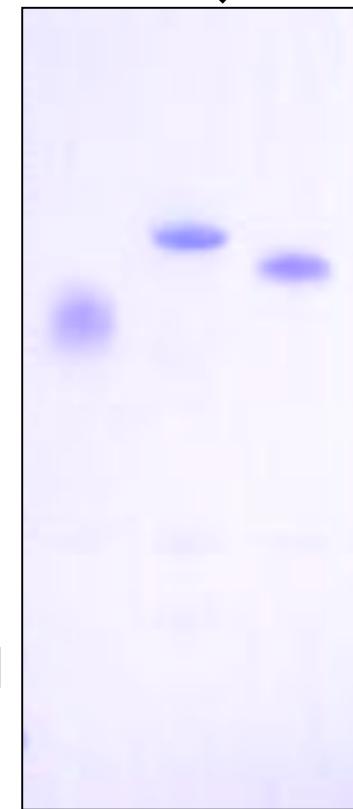


SDS PAGE

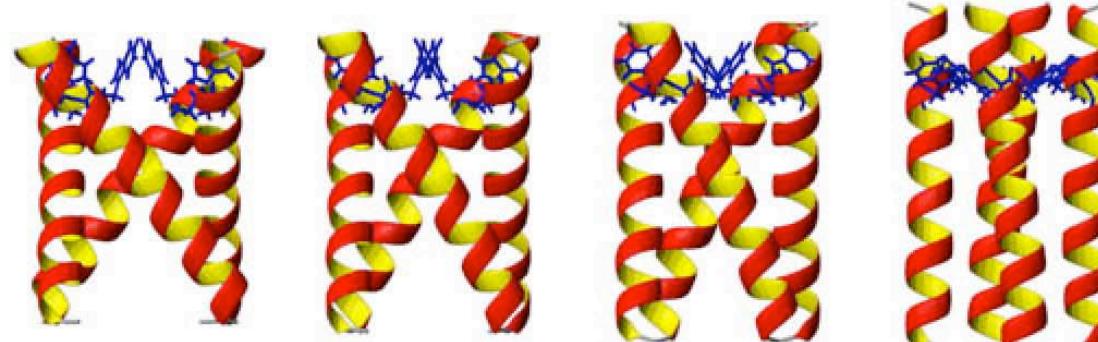
wild-type Vpu TM  
A18H Vpu TM



wild-type Vpu  
M2/Vpu chimera  
A18H Vpu



*Structure varies with the type of biological membrane.*

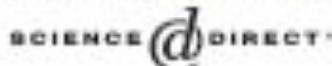
Type of biological membrane	Basolateral plasma membrane	Endoplasmic reticulum	Golgi	Apical plasma membrane
Hydrophobic thickness (A)	$23.1 \pm 0.6$	$25.0 \pm 0.4$	$27.0 \pm 0.4$	$30.0 \pm 0.3$
Predicted tilt angle (degrees)	40	34	26	4
Tetramer model				

# *A18H mutation makes Vpu of HIV-1 act like M2 of influenza.*

M2 TM domain	-SSDPLVVAASIIGIL <b>H</b> LIL <b>WILDRL</b> -
Wild Vpu2-30+	QPIQIAIVALVVAAIIIAIVV <b>WSIVIIEGRGGKKKK</b>
A18H Vpu2-30+	QPIQIAIVALVVAAII <b>HIVVWSIVIIEGRGGKKKK</b>



Available online at [www.sciencedirect.com](http://www.sciencedirect.com)



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Virology xx (2006) xxx–xxx

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VIROLOGY

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[www.sciencedirect.com/locate/yviro](http://www.sciencedirect.com/locate/yviro)

A single amino acid substitution within the transmembrane domain of the human immunodeficiency virus type 1 Vpu protein renders simian–human immunodeficiency virus (SHIV<sub>KU-1bMC33</sub>) susceptible to rimantadine

David R. Hout, Lisa M. Gomez, Erik Pacyniak, Jean-Marie Miller, M. Sarah Hill,  
Edward B. Stephens \*

*Department of Anatomy and Cell Biology, University of Kansas Medical Center 3901 Rainbow Blvd., Kansas City, KS 66160, USA*

*Received 7 September 2005; returned to author for revision 8 November 2005; accepted 9 December 2005*

## *A18H mutation makes changes structure of Vpu.*

M2 TM domain

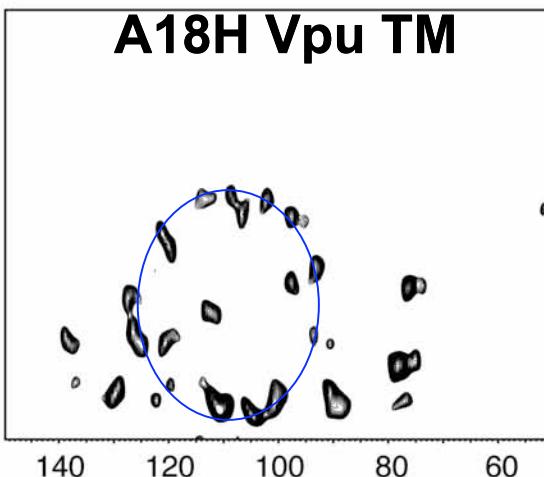
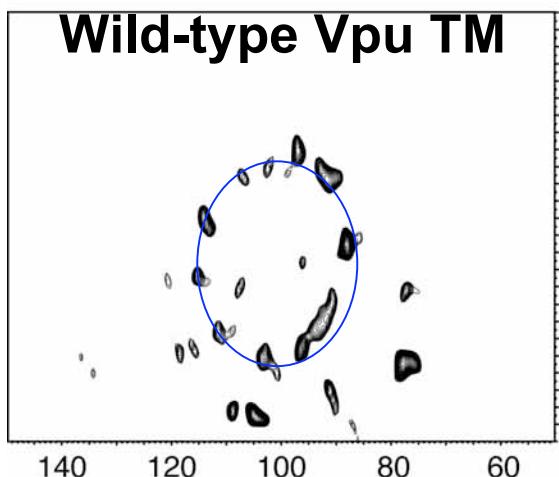
-SSDPLVVAASIIIGIL**H**LIL**WILDRL**-

Wild Vpu2-30+

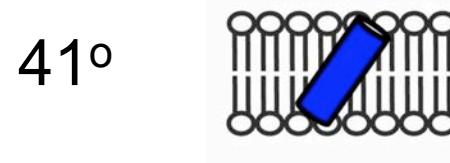
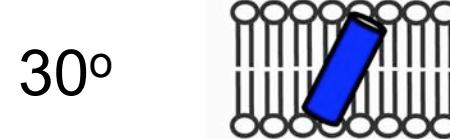
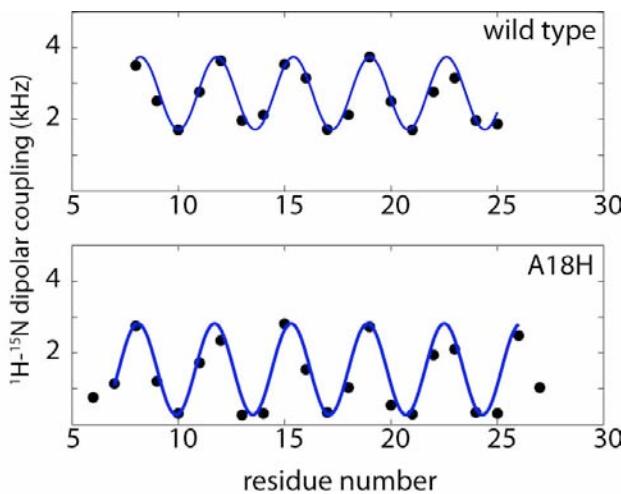
QPIQIAI~~VALVVA~~I~~III~~AI~~VV~~**WSIVI**IEGRGGKKKK

A18H Vpu2-30+

QPIQIAI~~VALVVA~~I~~III~~**HIVVWSIVI**IEGRGGKKKK



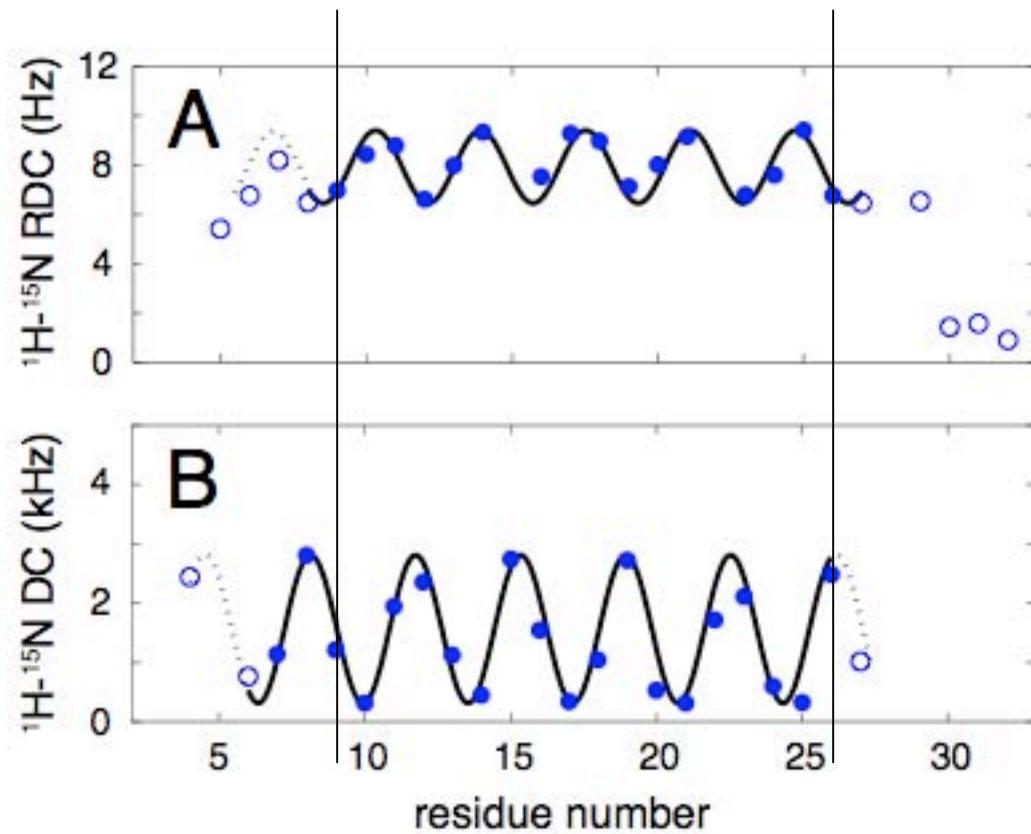
**Mutation lengthens helix and increases tilt angle.**



Vpu mutation: Hout et al 2006

## *A18H Vpu TM domain: Micelles vs. Bicelles.*

### Dipolar Waves

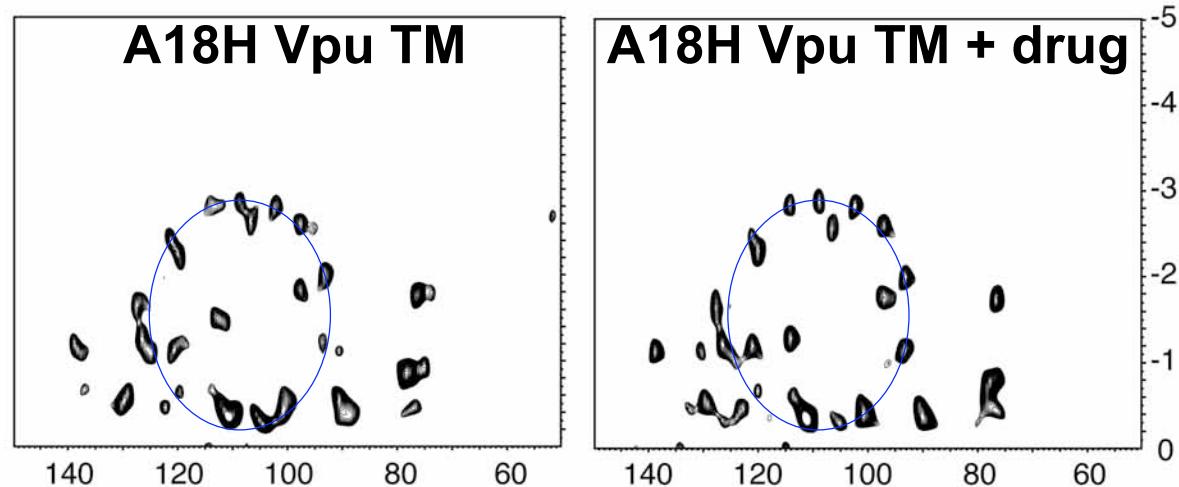


DHPC micelles  
 $q=0$

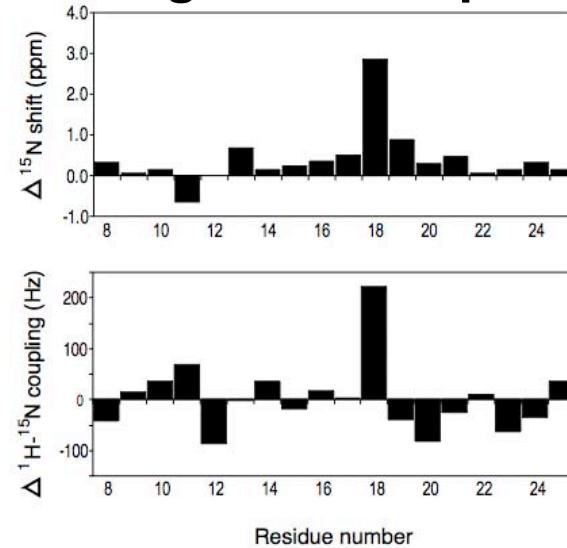
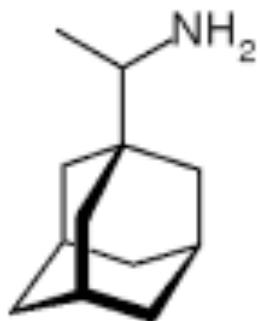
DMPC:DHPC bicelles  
 $q=3.2$

# *A18H mutation makes HIV-1 infection sensitive to rimantidine.*

M2 TM domain	-SSDPLVVAASIIIGIL <b>H</b> LIL <b>WILDRL</b> -
Wild Vpu2-30+	QPIQIAI <del>VALVV</del> AIII <del>IAIVV</del> <b>WSIVI</b> IEGRGGKKKK
A18H Vpu2-30+	QPIQIAI <del>VALVV</del> AIII <b>HIVV</b> <b>WSIVI</b> IEGRGGKKKK

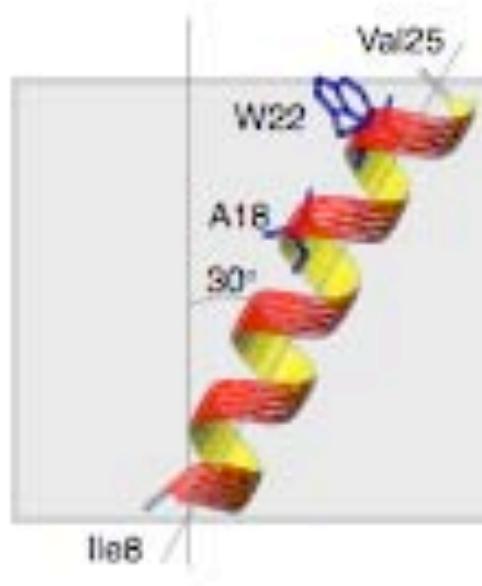


Drug binding induces spectral changes

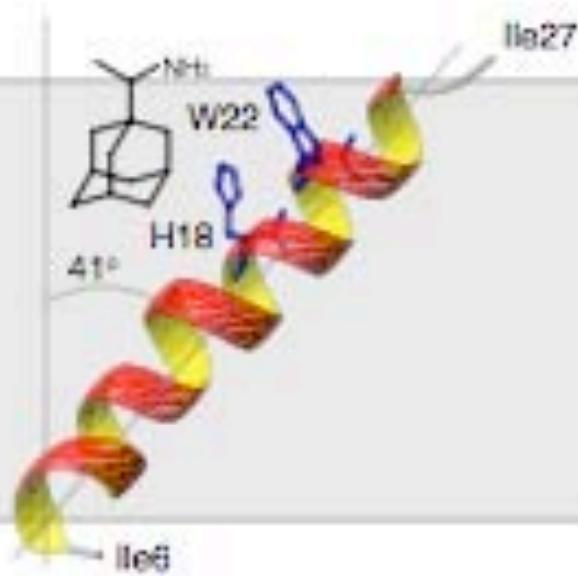


## *Structures of Vpu TM: wild-type vs. A18H mutant.*

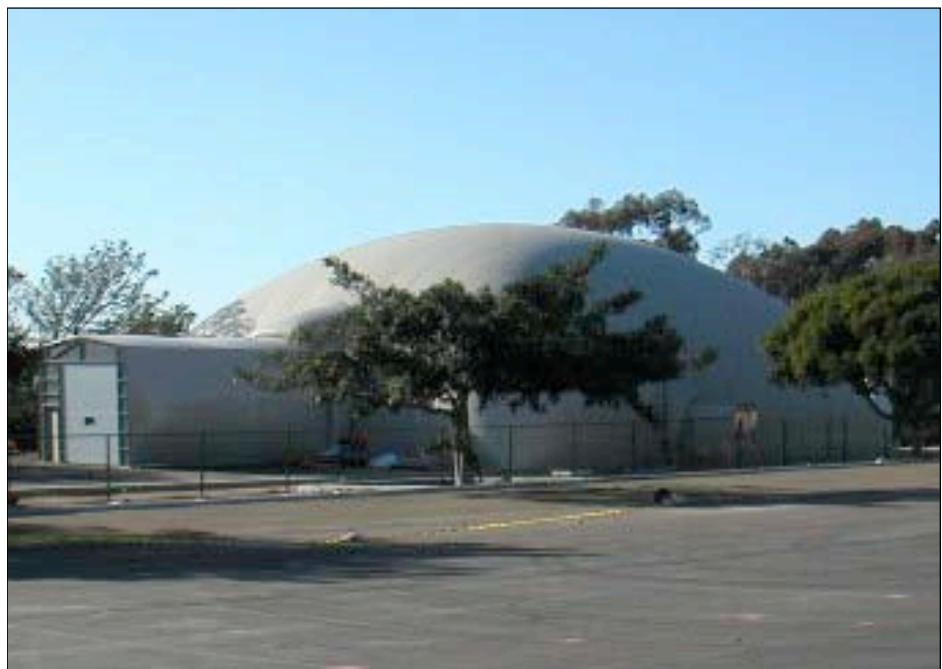
**Wild type Vpu TM**



**A18H Vpu TM + drug**



# *Biotechnology Resource for NMR Molecular Imaging of Proteins at UCSD.*



Supported by the National Institutes of Health



## Mer project

Anna DeAngelis  
Stanley Howell  
Annie Pham  
\*Woo Sung Son

## Viroporin project

\*Gabriel Cook  
Fabian Filipp  
Sang Ho Park  
\*Yan Wang

## Solid-state NMR

Chris Grant  
Alex Nevzorov  
Neeraj Sinha  
Albert Wu

## Coat proteins and lipids

\*David Black

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Francesca Marassi (Burnham Institute)  
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\*Maha Radhakrishnan